

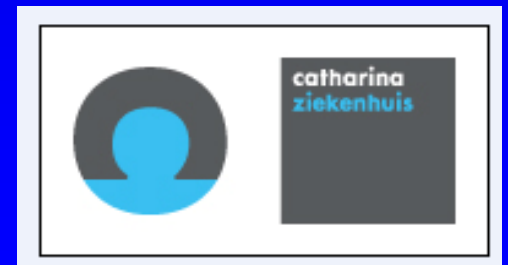
***CORONARY PHYSIOLOGY IN THE CATHLAB:***

**ABSOLUTE BLOOD FLOW MEASUREMENTS:  
*PRINCIPLES***

***Educational Training Program ESC  
European Heart House  
april 23rd - 25th 2015***



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# PROBLEMS WITH ABSOLUTE FLOW (and FLOW VELOCITY)

- **absolute blood flow** has no meaning without knowledge of the extent of the perfusion area or without knowing a normal value
- absolute flow and all **flow-derived parameters** are dependant on perfusion pressure, which is highly variable within the same patient
- **coronary** blood flow is often not representative for **myocardial** perfusion, especially not in case of severe stenosis
- coronary flow, velocity, and CFR do not discriminate **epicardial or microvascular disease**

Consequently, for *routine interventional practice* and decision making in the catheterization lab, absolute flow or flow velocity has little value.

*Fractional Flow Reserve (FFR)* perfectly describes the influence of the epicardial stenosis on myocardial perfusion

For *scientific purposes* and assessing the *microcirculation*, things might be different

***If you know absolute maximum blood flow***

in ml/min for an arbitrary stenosis, arbitrary myocardial distribution and any blood pressure and heart rate  
(e.g. 80 ml / min)

**AND you know FFR**

(e.g. 0.50)

***you also know the normal maximum flow*** for that respective distribution under those specific hemodynamic conditions:

→ ***160 ml / min (!)***

.....and if you also know **coronary wedge pressure ( $P_w$ )**, you know both the **absolute myocardial, coronary, and collateral flow**

*(because FFR gives the relative contribution of coronary arterial and collateral flow to myocardial flow)*

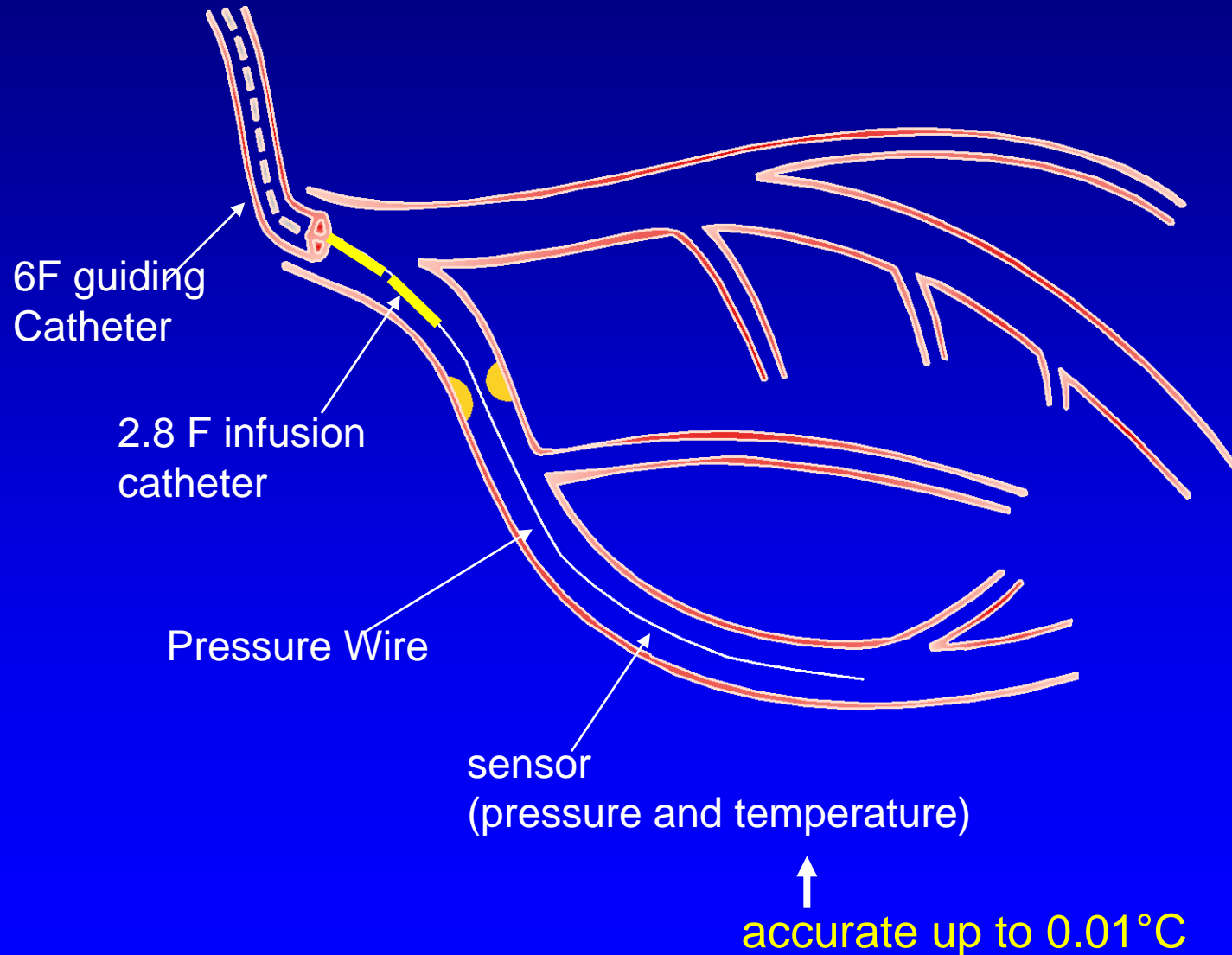
.....and if you measure **coronary pressure** simultaneously, you know **all resistances quantitatively** for any arbitrary coronary segment, myocardial distribution, or collateral bed

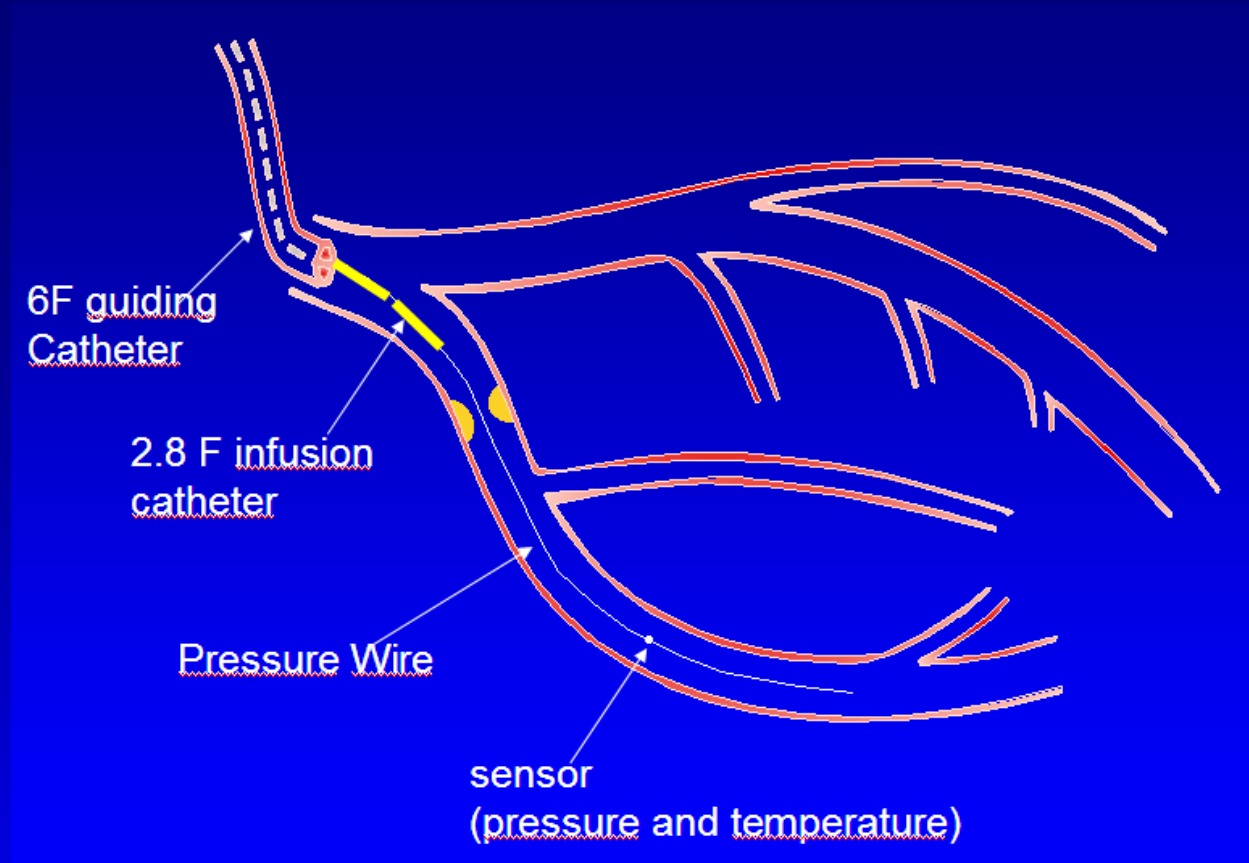
————→ **Easy assessment of microcirculation and collaterals**

***So, from the scientific and physiologic point of view, it would be attractive to measure hyperemic absolute flow (in conjunction with pressure)...***

- assessing the microcirculation
- quantitation of collateral flow
- quantitation of microvascular flow and resistance in specific patient groups:
  - *heart-transplant follow-up*
  - *stem cell therapy*
  - *syndrome X (whatever it might be)*
  - *prognostic stratification after STEMI*

# continuous infusion of saline to determine absolute maximum coronary blood flow





saline infused at 20 ml/min

temperature of saline is 5° below blood temperature

after mixing, temperature of mixtate is 1° below blood temp

—————> blood flow must be 5 x infusion flow of saline



Absolute  
max coronary  
blood flow



infusion flow



$$Q_b = Q_i \cdot \frac{T_i}{T} \cdot 1.08$$

temp of infusion at tip  
of guiding catheter

temp of blood at  
sensor position

saline infused at 20 ml/min

temperature of saline is 5° below blood temperature

after mixing, temperature of mixtate is 1° below blood temp

—————> blood flow must be 5 x infusion flow of saline

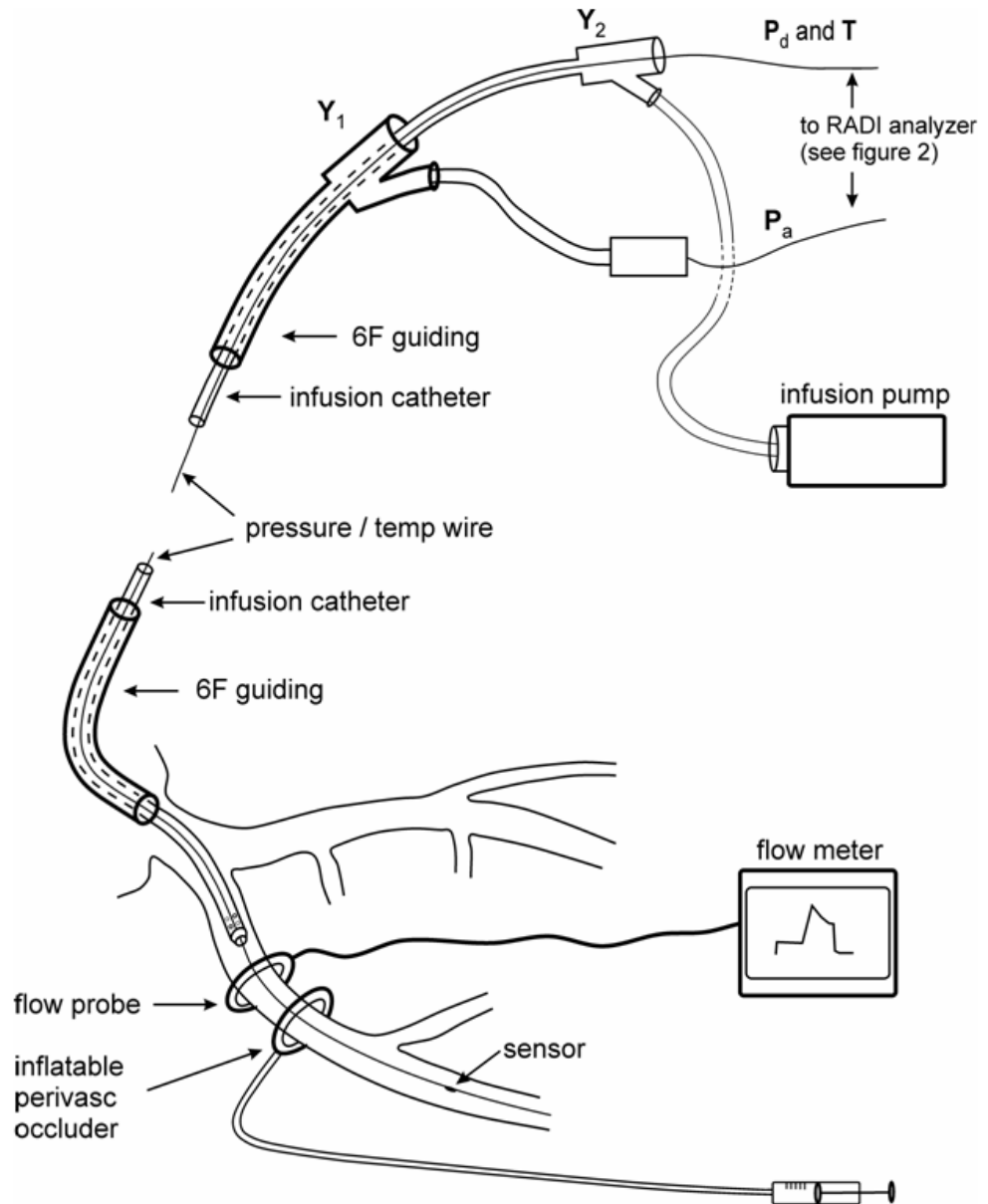
# continuous infusion of saline to determine absolute maximum coronary blood flow

## Prerequisites :

- decrease of coronary temperature should be large enough to be detected and stable ( or stated another way: temperature sensor should be sensitive enough to detect changes) , given a reasonable injectate flow ( *adequate signal/noise ratio* )
- *complete mixing* of blood and saline between injection site ( = tip of infusion catheter) and sensor position
- *heat ( “cold” ) loss through the wall* of the artery must be small compared to convection by blood

# Animal study: instrumentation

Figure 1



# ***ANIMAL STUDY: methods***

- five mongrel dogs, 28-43 kg
- instrumentation of LCX by perivascular flow probe and perivascular balloon occluder at day 0
- cardiac catheterization at day 7, Amplatz guiding catheter
- 0.014" pressure wire (*RADI*) distal to flow probe/occluder for coronary pressure & temperature measurement
- 2.8 F special infusion catheter (*AMT, Inc*) over wire, position just proximal to occluder
- **creation of different degrees of stenosis**  
*(corresponding with FFR of 0.85, 0.70, 0.55, 0.40)*
- measurements with continuous saline infusion at 2 different infusion rates and 2 different sensor positions
- in-duplo measurements for all degrees of stenosis, both infusion rates, and both sensor positions:

**→  $4 \times 2 \times 2 \times 2 = 32$  measurements per dog**

# **Infusion Catheter For Thermodilution (Hexacath R)**

(complete mixing of blood and saline)



without guidewire



with guidewire

PRINT

EDIT

RENAME

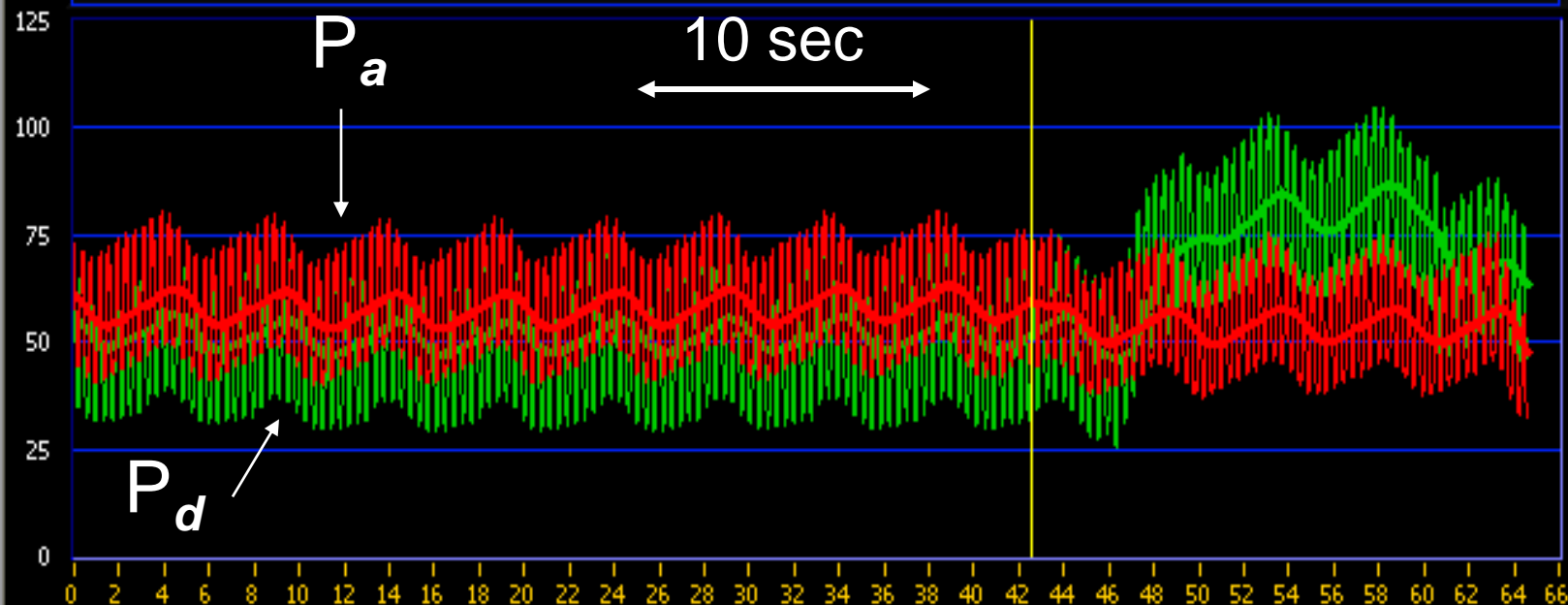
EXPORT

ERASE

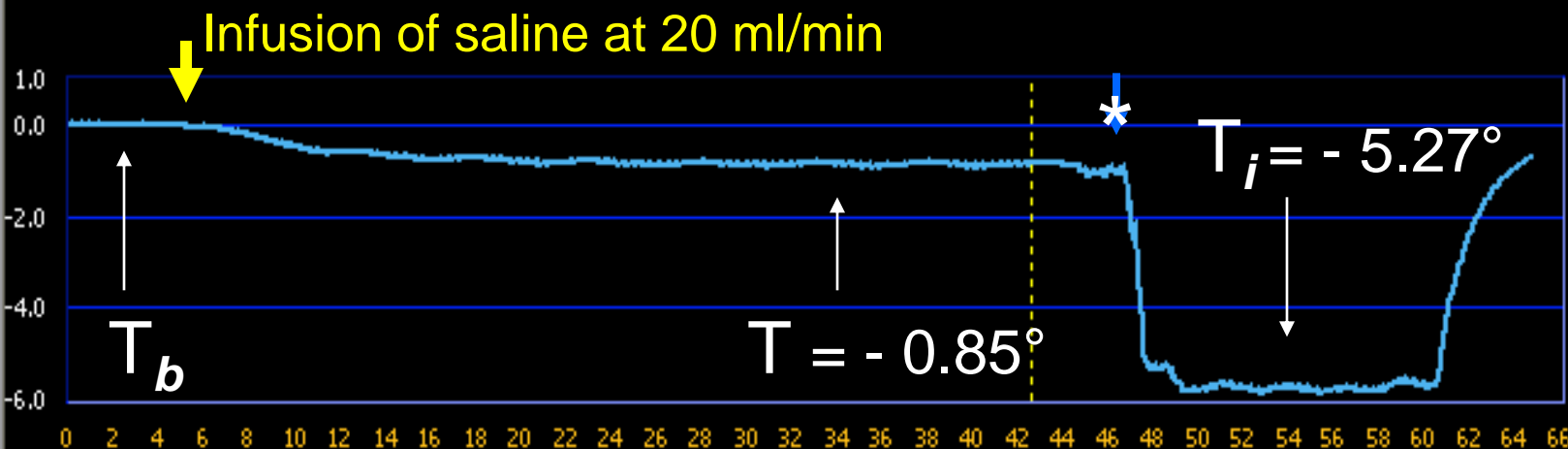
SETUP

Dog #5

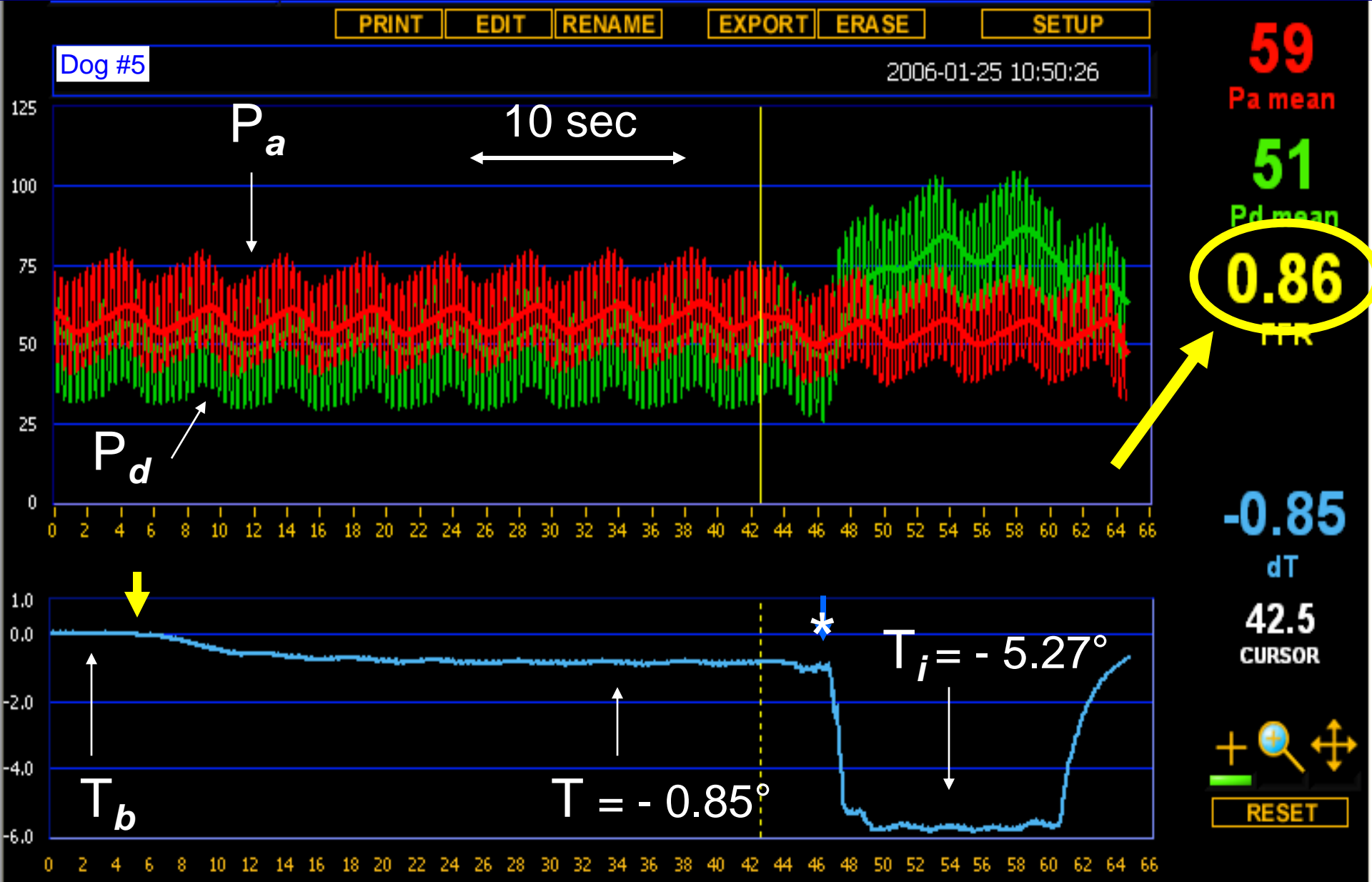
2006-01-25 10:50:26

**59**  
Pa mean**51**  
Pd mean**0.86**  
FFR**-0.85**  
dT**42.5**  
CURSOR

RESET

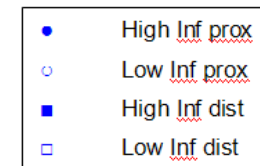
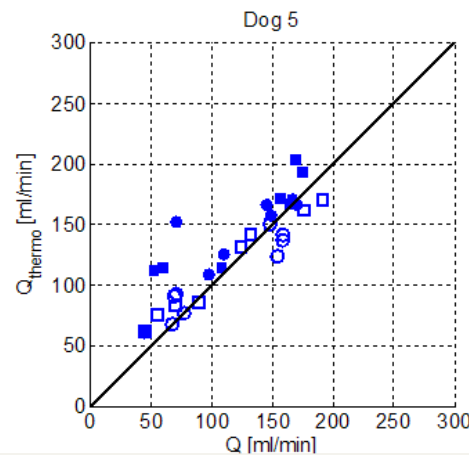
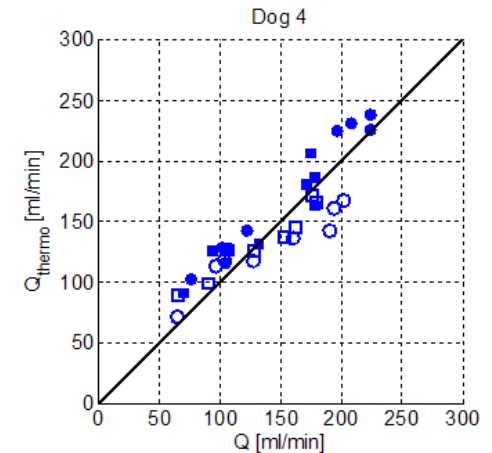
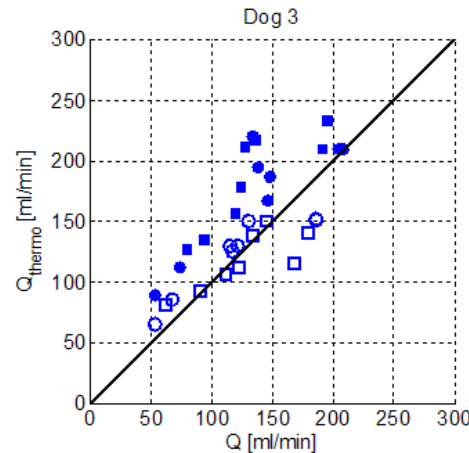
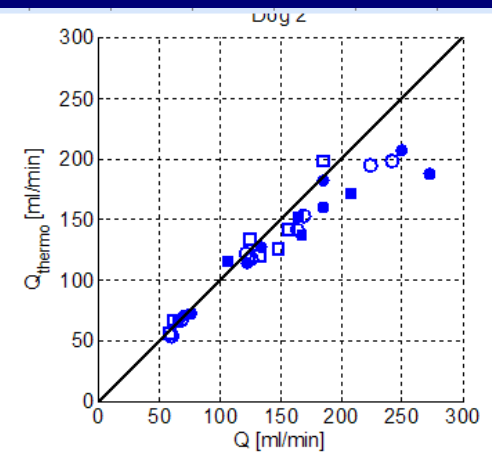
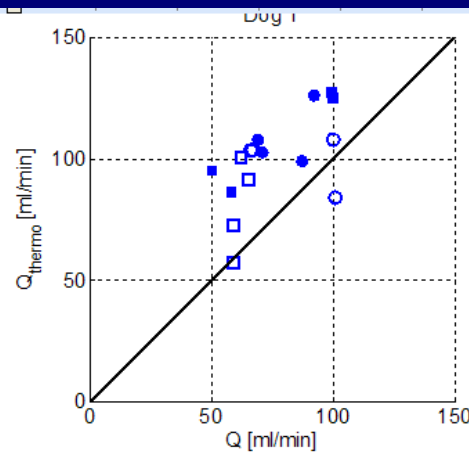


$$Qb = 20 \times (-5.27 / -0.85) \times 1.08 = 134 \text{ ml/min}$$



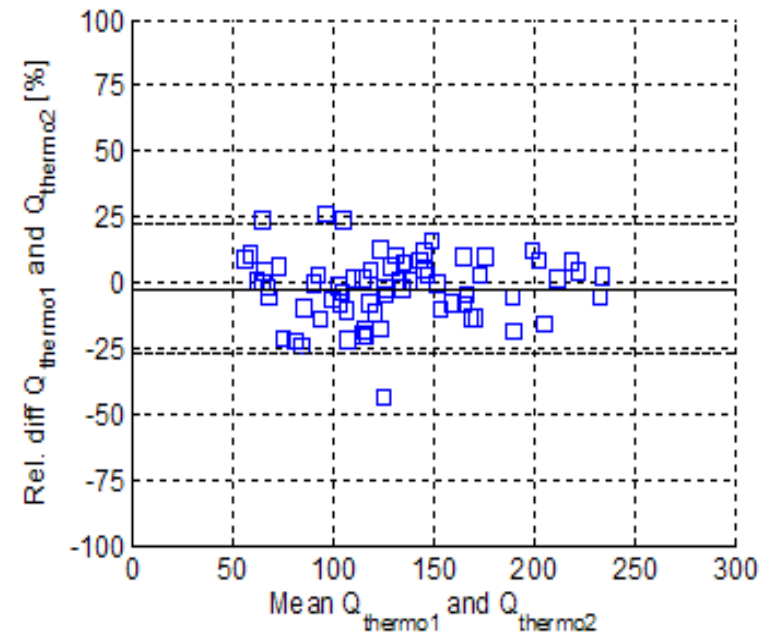
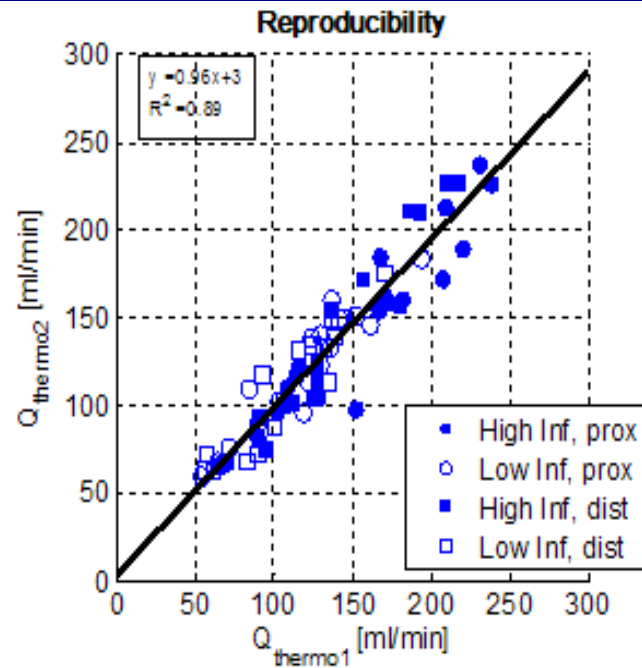
$Q_b = 134 \text{ ml/min} \rightarrow \text{normal max flow} = 100/86 \times 134 = 156 \text{ ml/min}$

# Animal study: results in the individual dogs



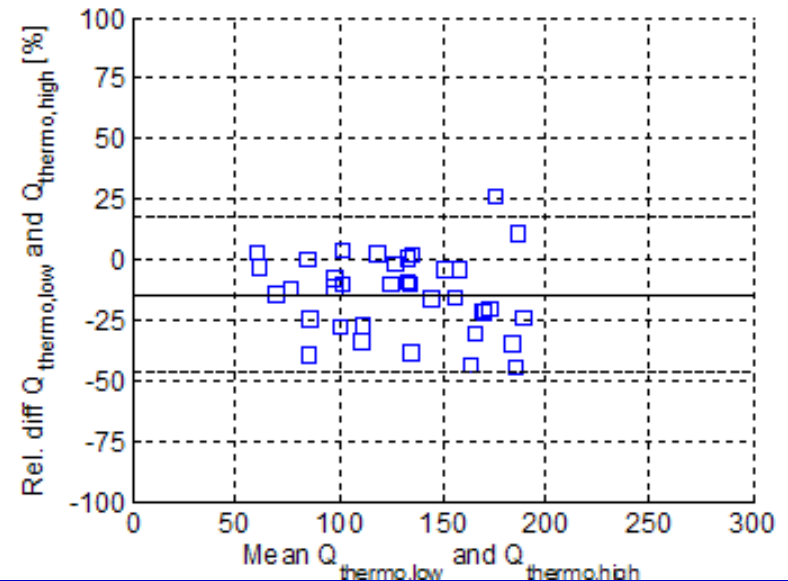
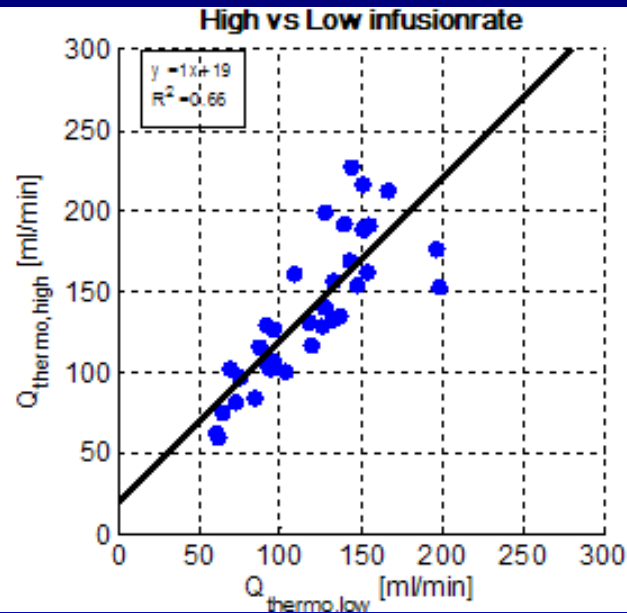


**b**



Animal study: *reproducibility* (N=72)

**C**

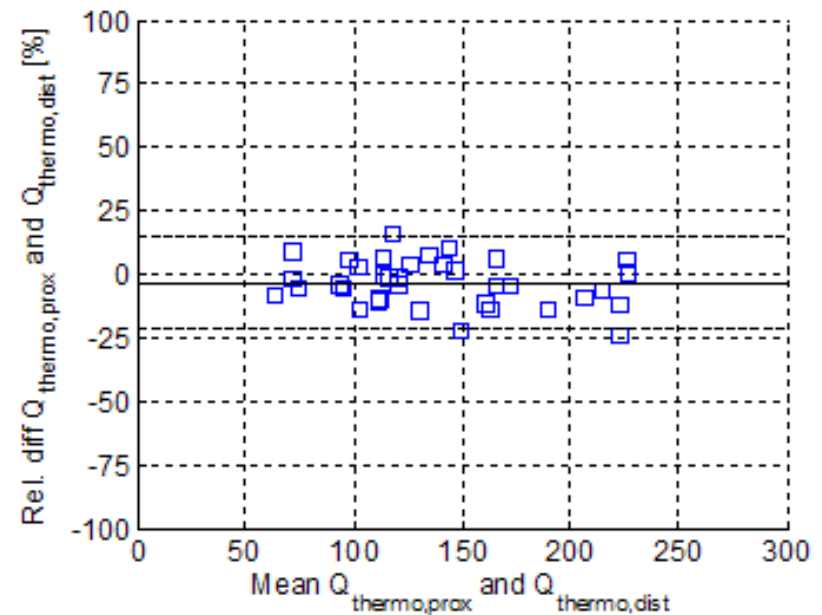
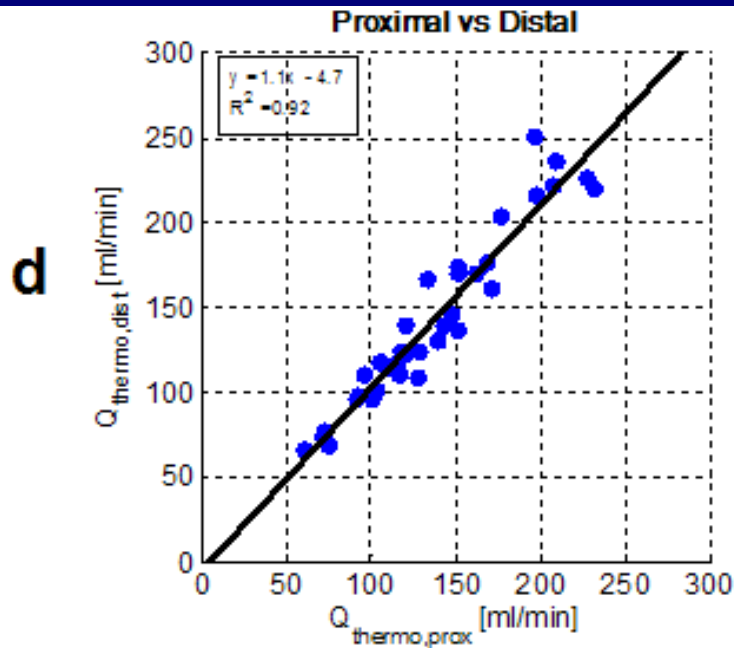


Animal study: *high versus low infusion rate*

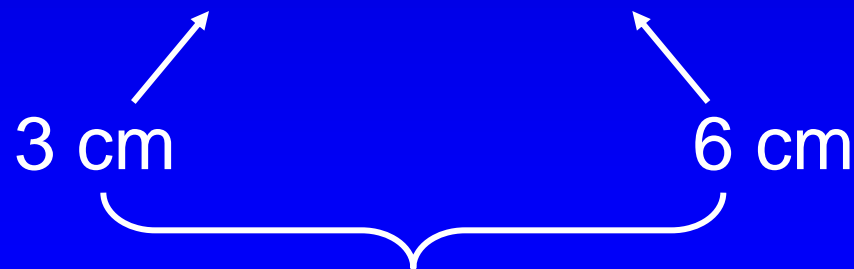
15-25 ml/min

8-15 ml/min

saline room temp



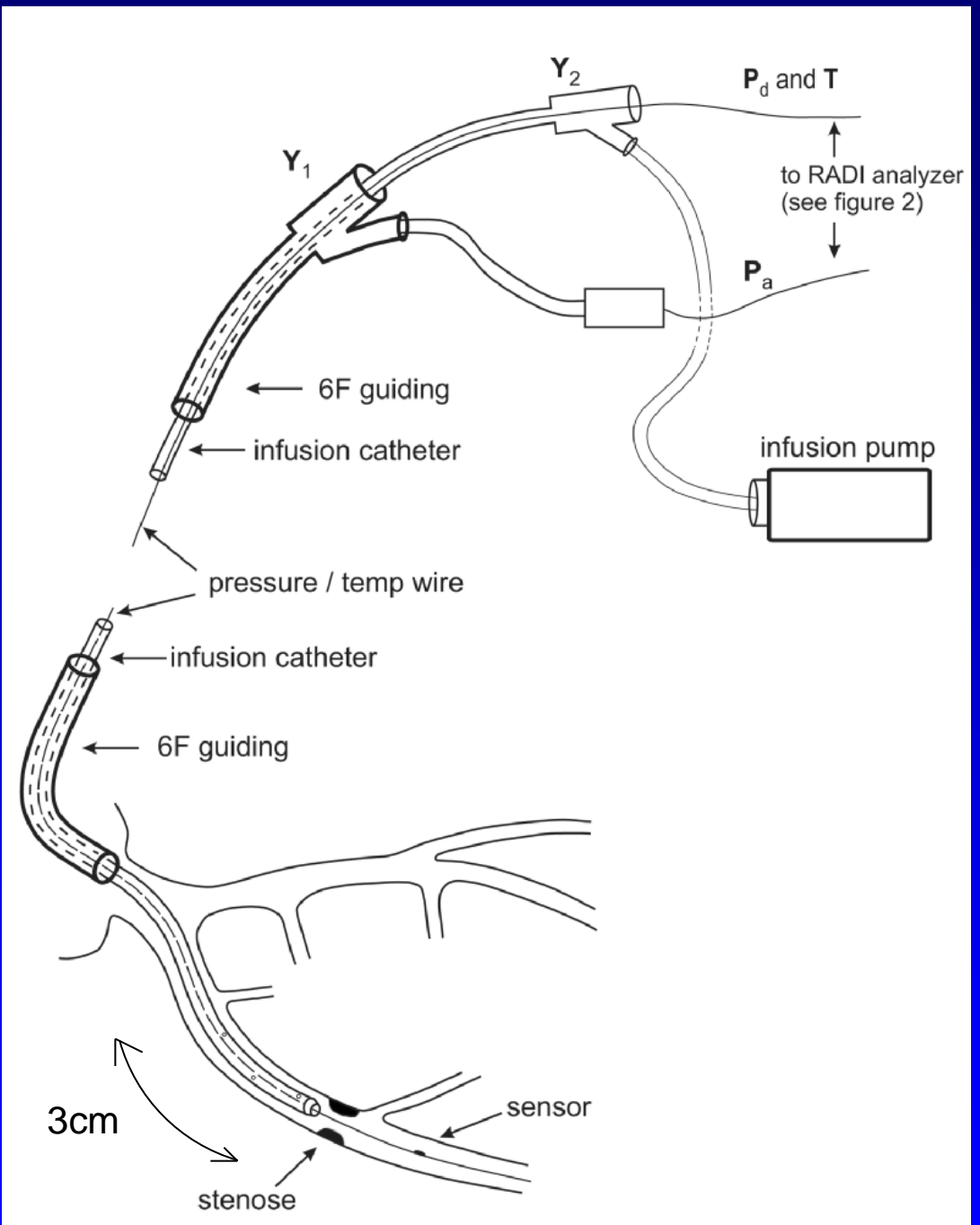
Animal study: *proximal versus distal sensor position*



from tip of infusion catheter

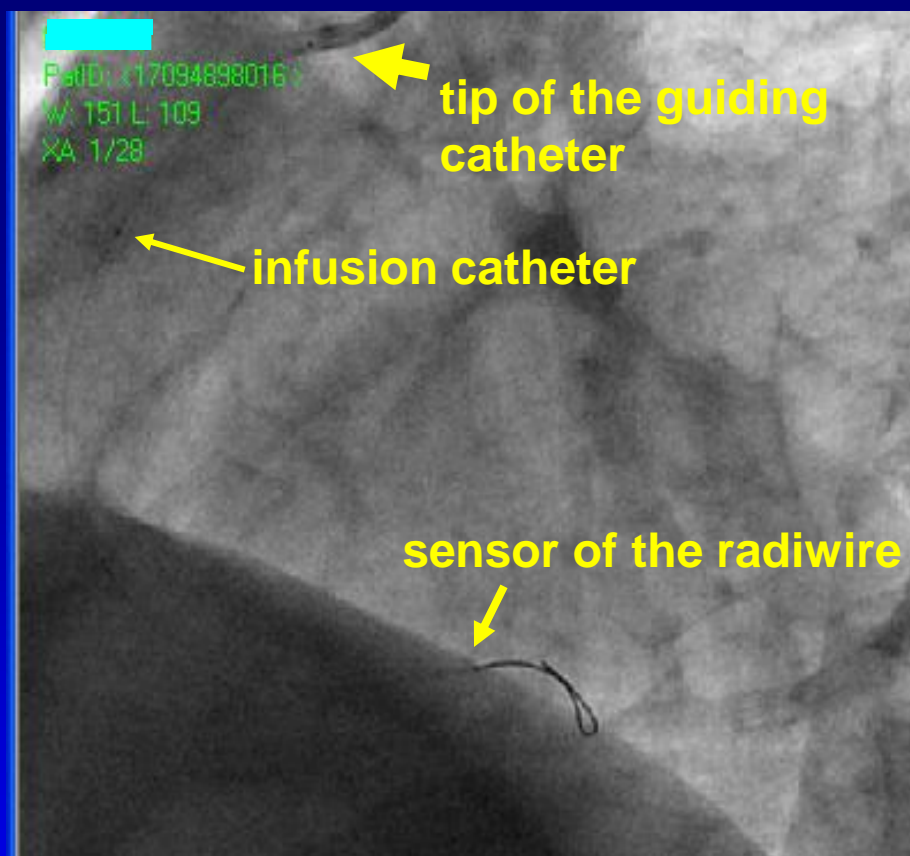
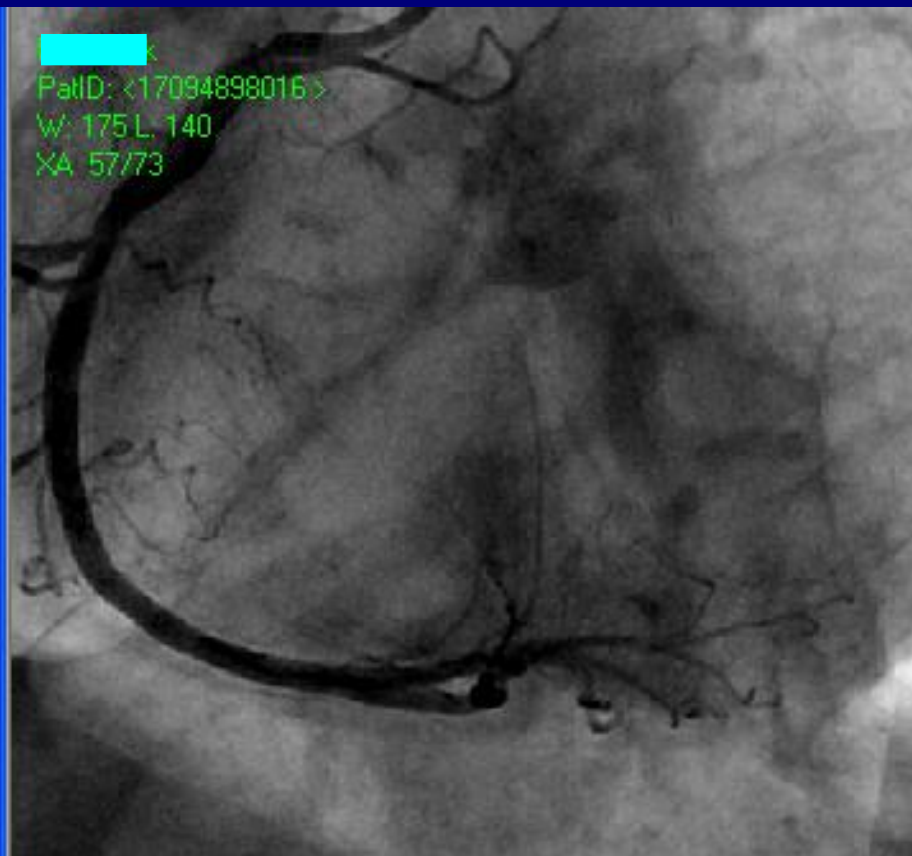
# Human validation study

# Human study: instrumentation

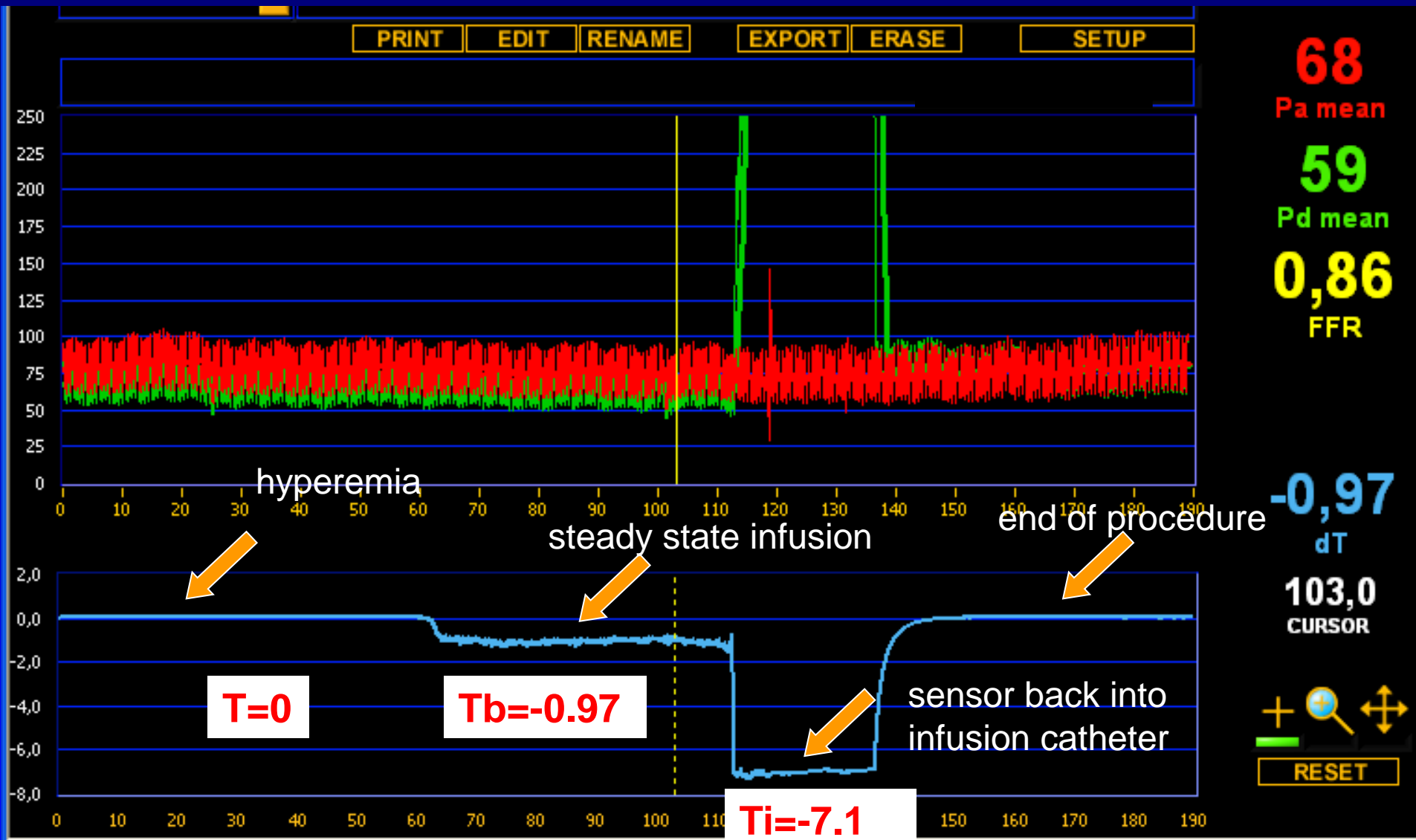


## Patient # 1:

- normal right coronary artery
- $Q_i = 25 \text{ ml / min}$  saline at room temperature
- sensor located 7 cm from tip of infusion catheter



# continuous infusion during 3 minutes



$$Q_b = 25 \times (7.1 / 0.97) \times 1.08 = 198 \text{ ml/min}$$

(and normal max flow in this artery is  $100/86 \times 198 = 230 \text{ ml/min}$ )



## **Patient # 3:**

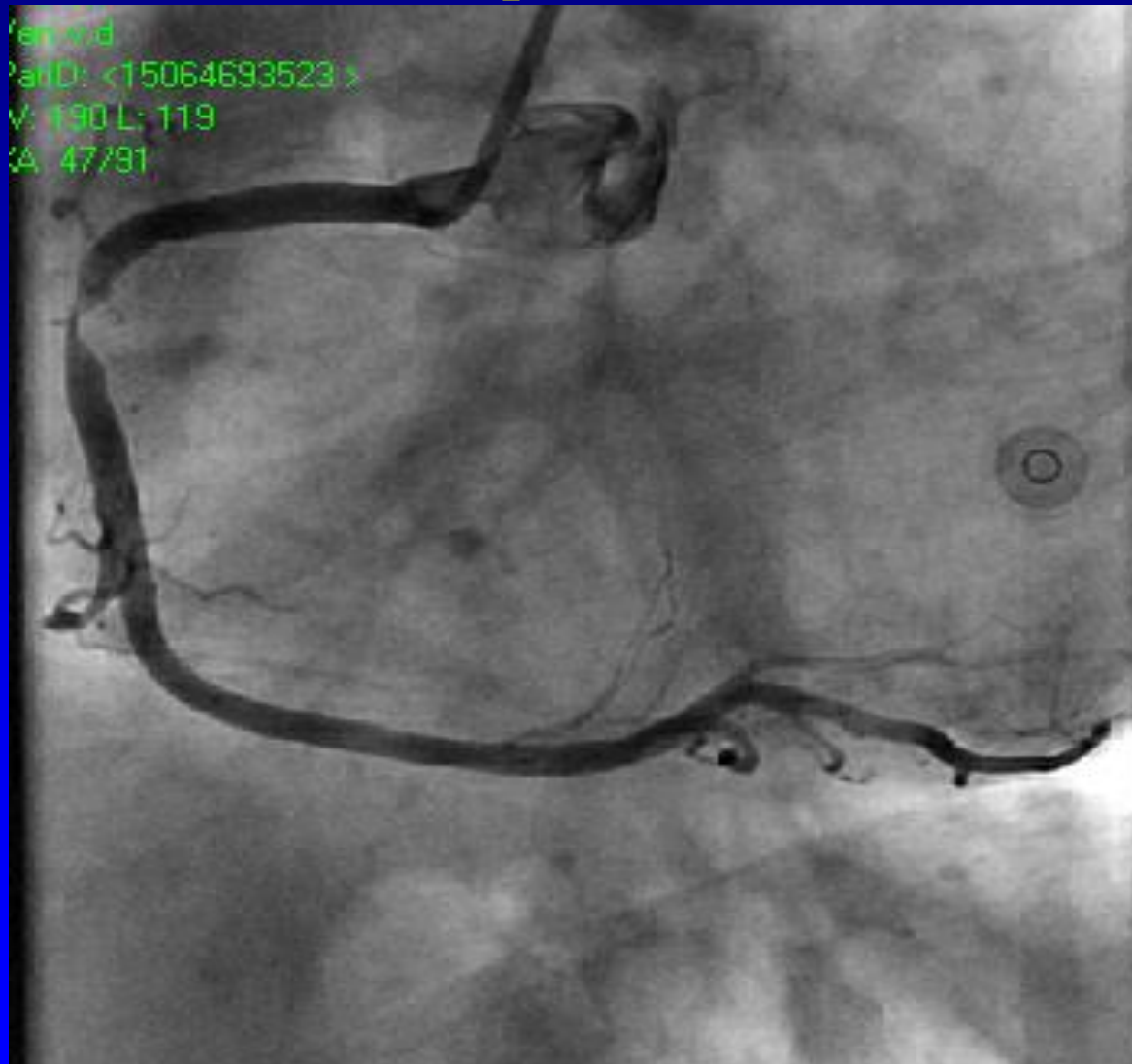
### **Before intervention**

- stenotic right coronary artery
- $Q_i = 15 \text{ ml / min}$  saline at room temperature
- distance 7 cm

### **After successful stenting:**

- $Q_i = 15 \text{ ml/min}$
- $Q_i$  is 20 ml/min

## Stenose proximale RCA

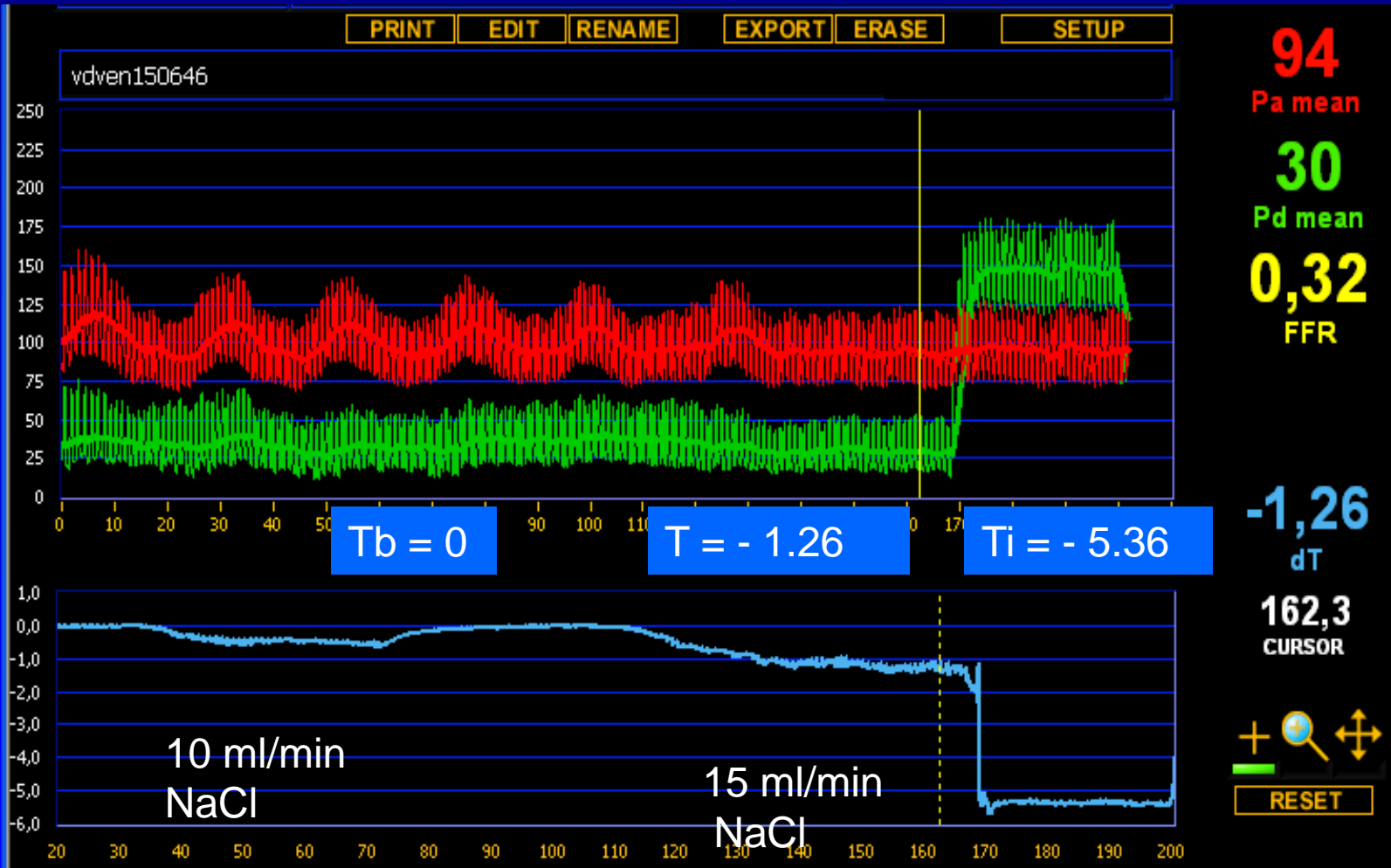


# FFR RCA= 0,42



# Infusion in RCA

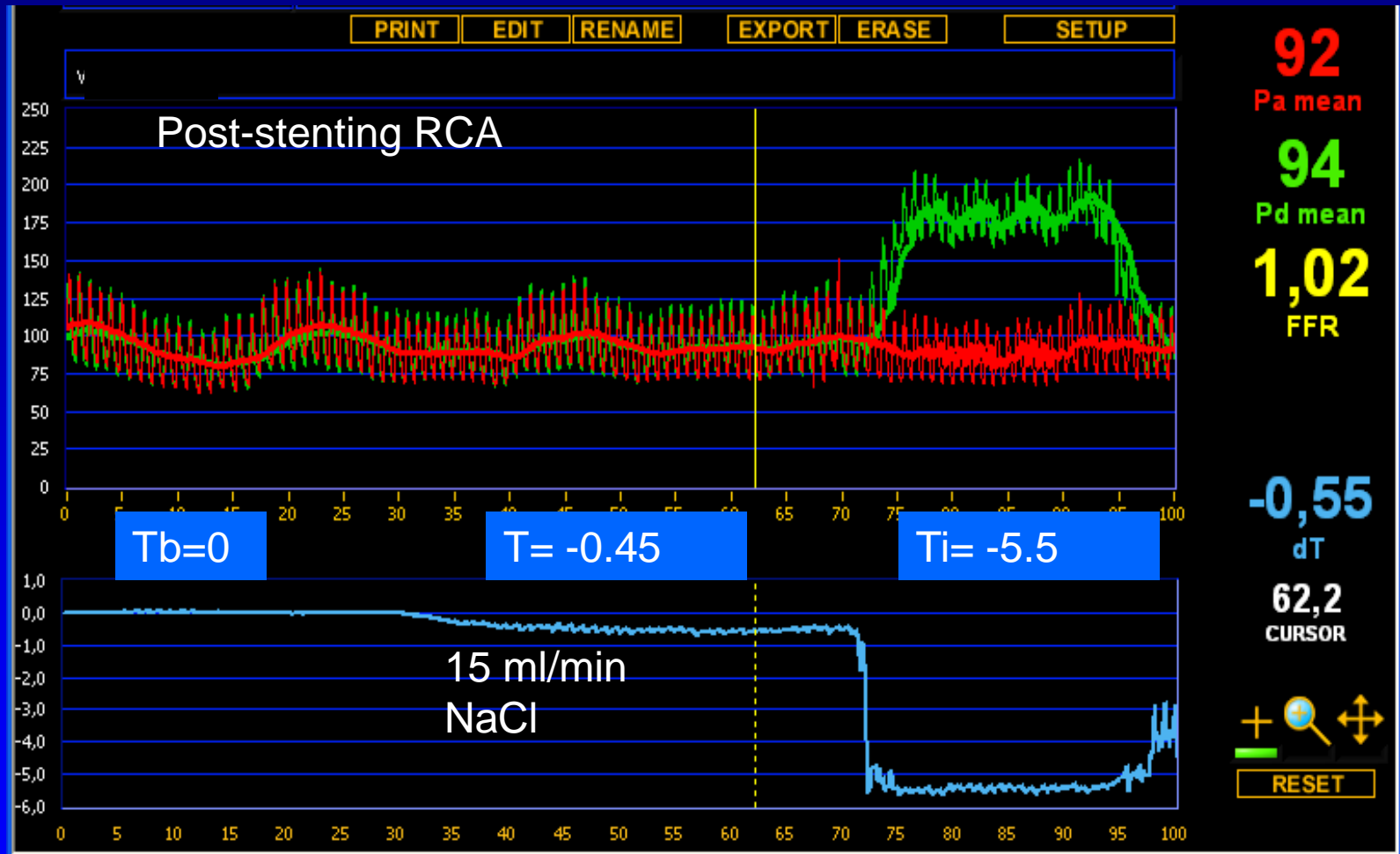
At first,  $Q_i=10\text{ml/min}$ , next infusion interrupted and restarted with a rate of  $Q_i=15\text{ml/min}$ .



$$Q_b = 15 \times (5.21 / 1.31) \times 1.08 = 64 \text{ ml/min}$$

# After stenting of RCA: FFR=0.98

Infusion rate  $Q_i = 15 \text{ ml/min}$ .



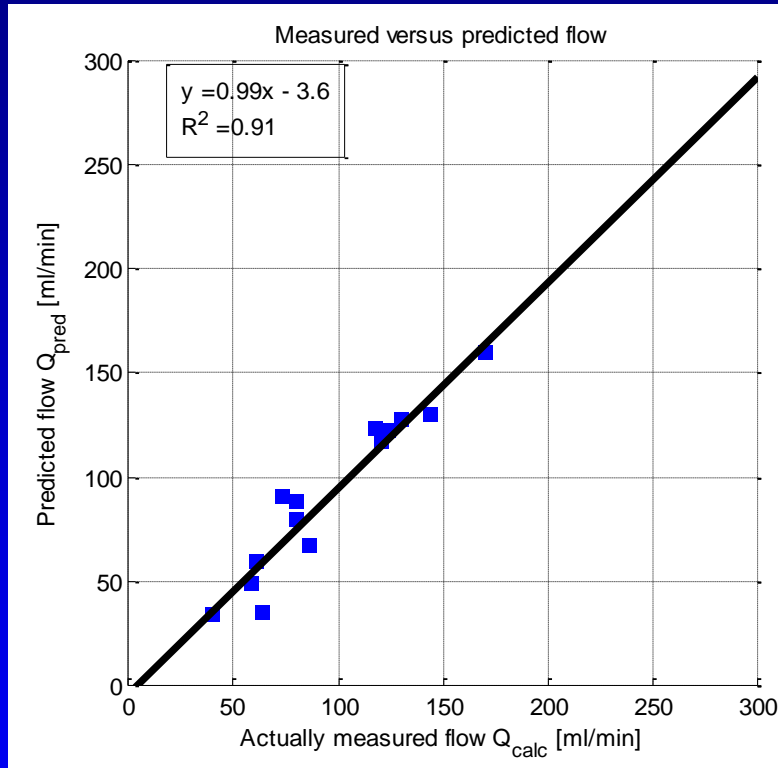
$$Q_b = 15 \times (5.5 / 0.45) \times 1.08 = 196 \text{ ml/min}$$

# ***Human Study: Selected Patients***

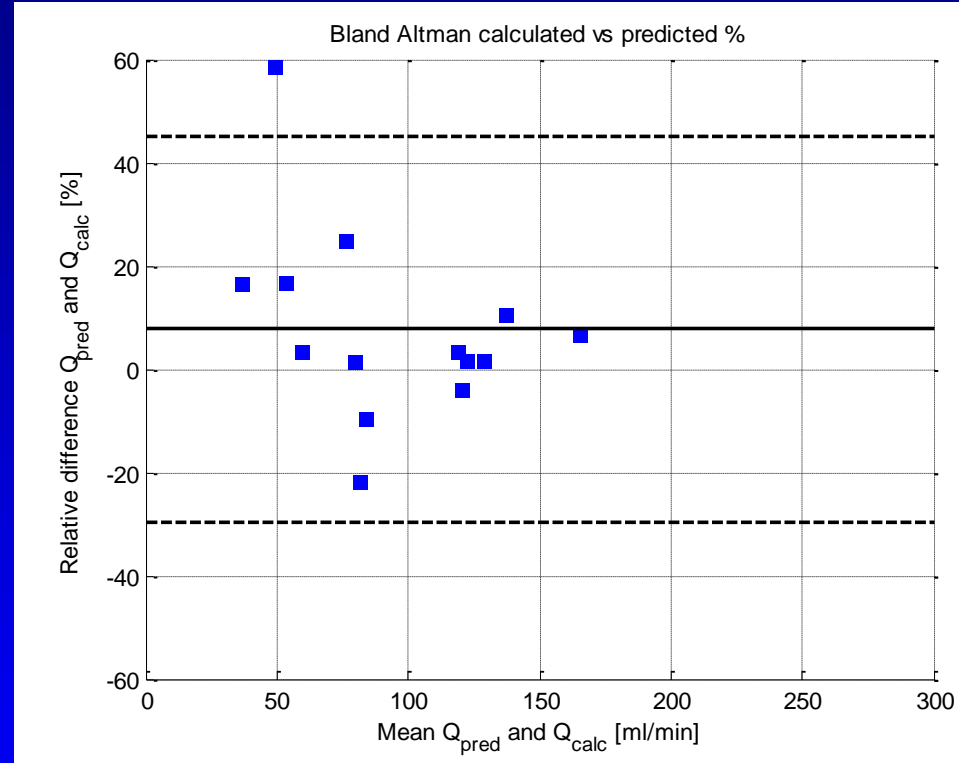
- 35 patients referred for PCI or FFR measurement
- single stenosis in segment without major sidebranches ( 24 RCA, 10 LAD, 1 LCX)
- $\text{FFR} \leq 0.75$  in 14 patients → stenting
- in these patients thermo measurement before and after PCI with saline at room temperature, for indirect quantitative validation
- in 10 patients 2 different infusion rates of saline (10-15 and 15-25 ml/min)
- in 11 patients 2 different sensor positions ( 3-4 and 6-8 cm distal to the tip of the infusion catheter)
- all measurement in duplo with 2-min interval in between

# Human study: quantitative validation

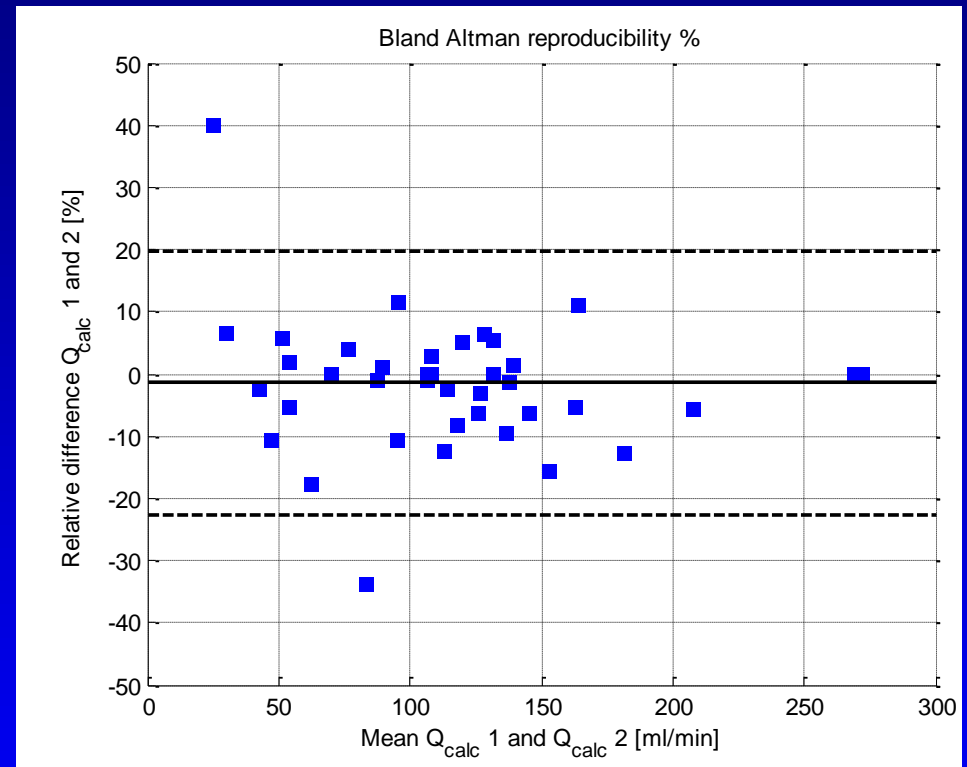
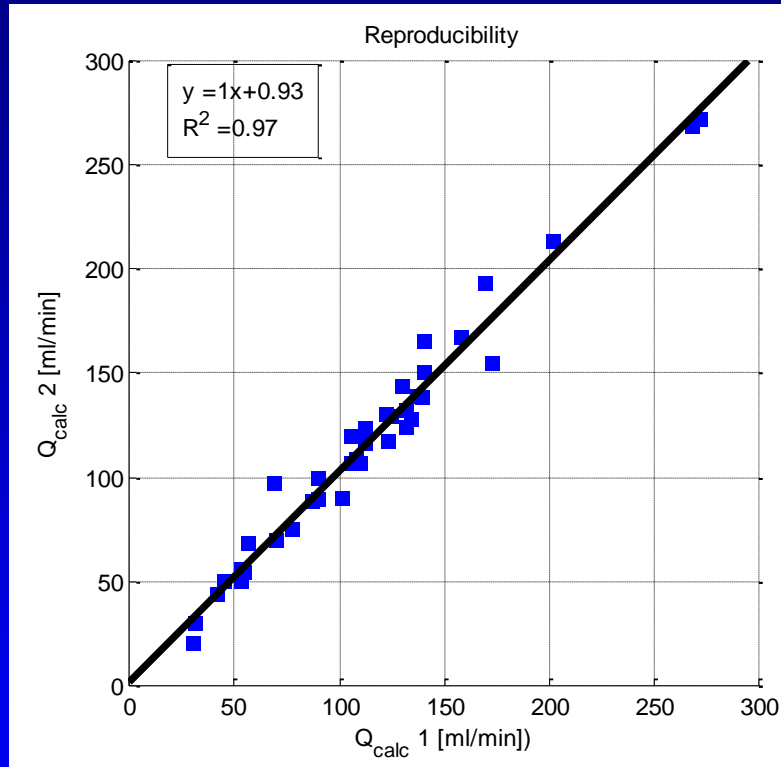
predicted flow



measured flow



# Human study: reproducibility





Clinical application:

Absolute Myocardial Flow and  
Microvascular Resistance in  
Acute Myocardial Infarction  
and at Follow-up

*Wijnbergen et al, submitted*

absolute flow  
in the infarct  
area  
(ml/min/g)

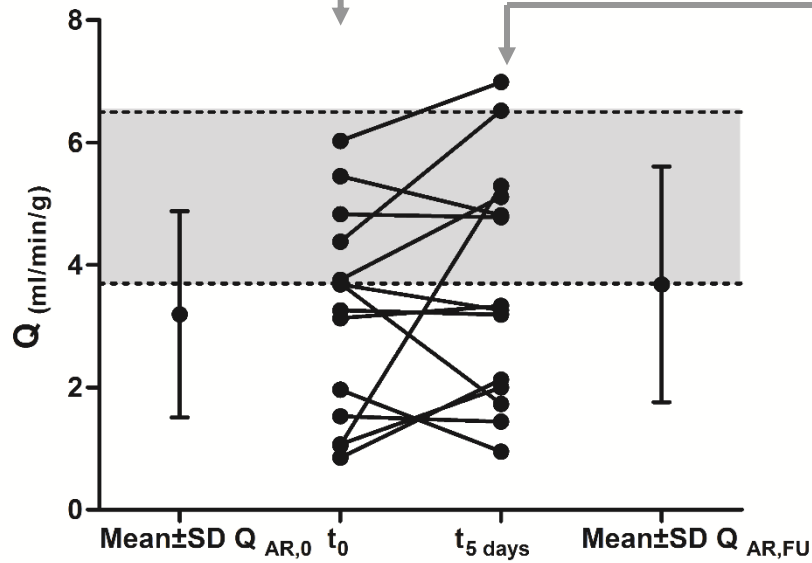


Figure 4a

absolute  
resistance  
in the infarct  
Area  
(dyn.s.cm<sup>-5</sup>)

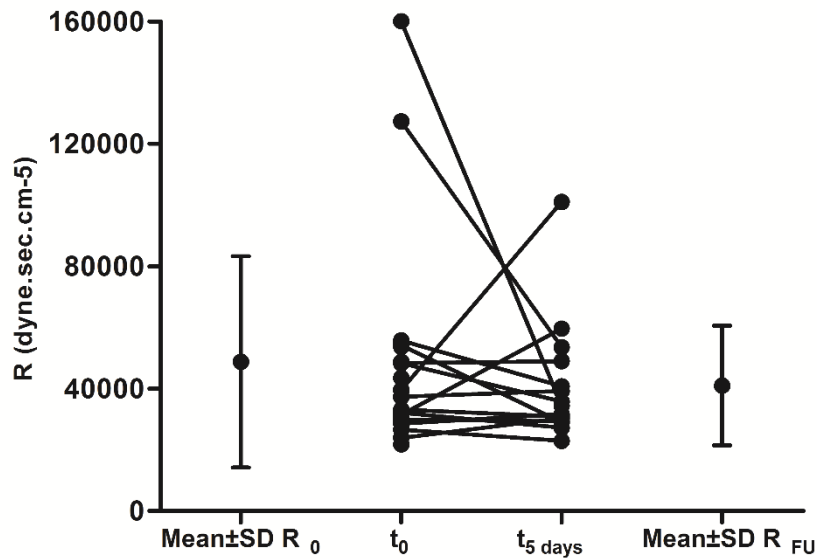


Figure 4b

hyperacute phase  
day 5

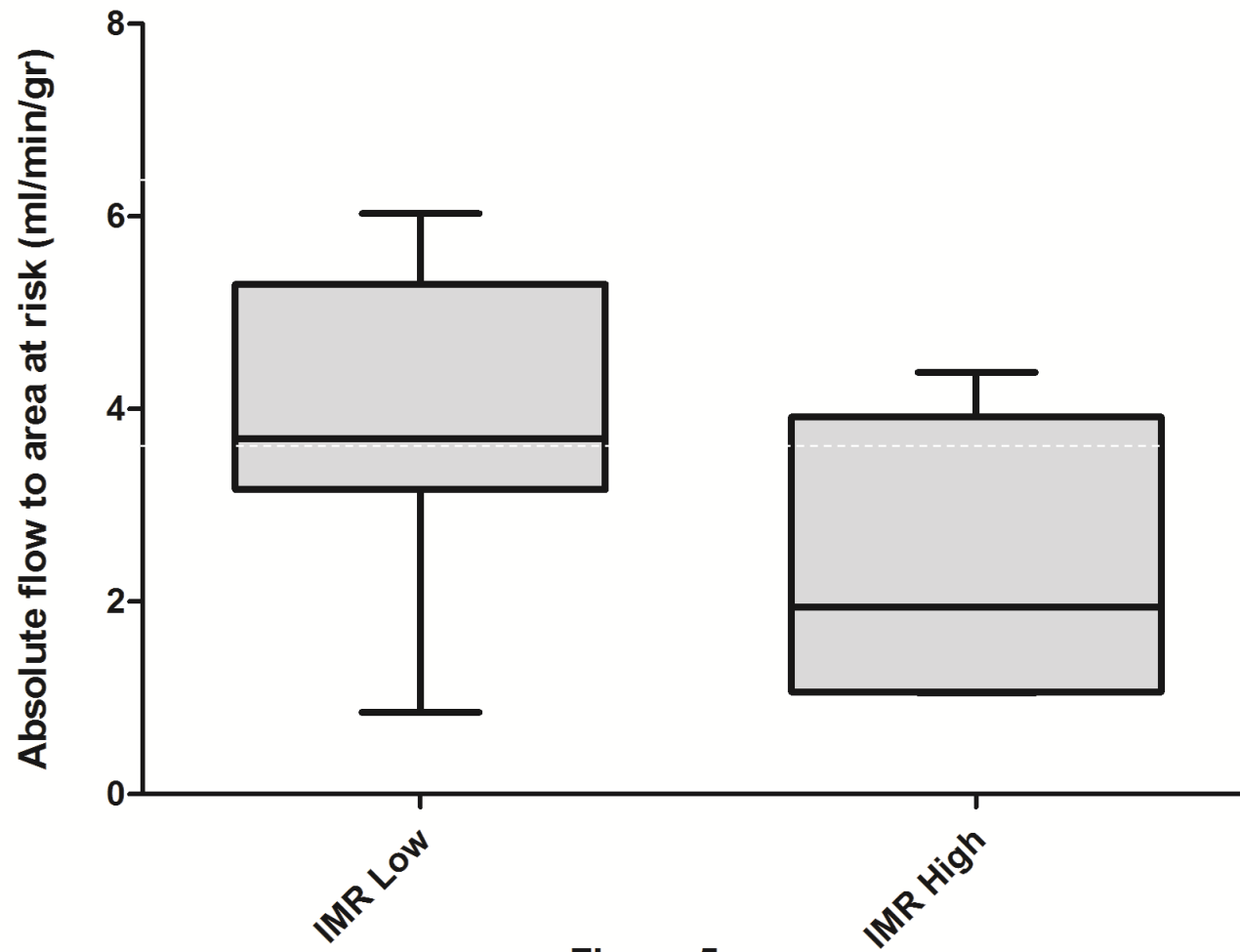
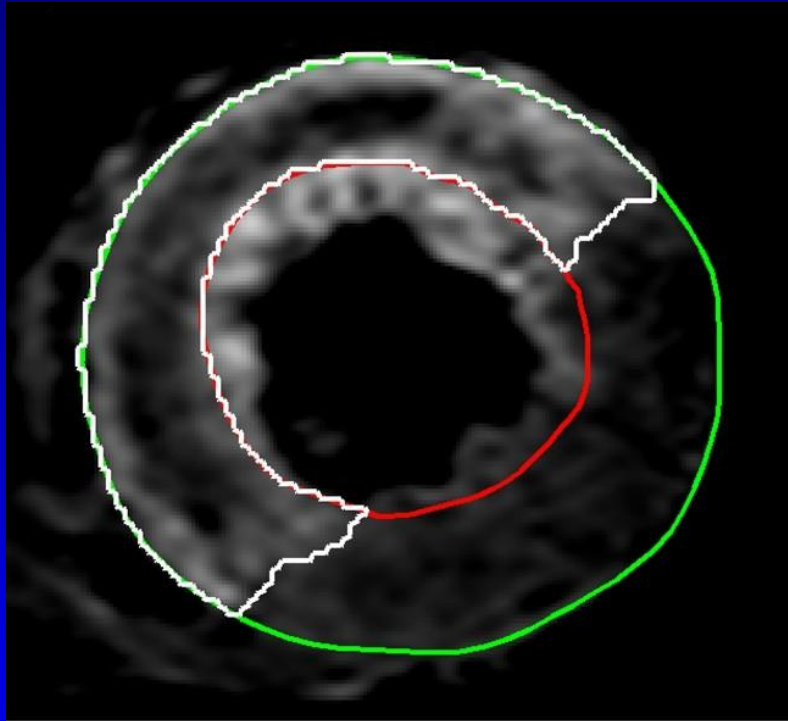
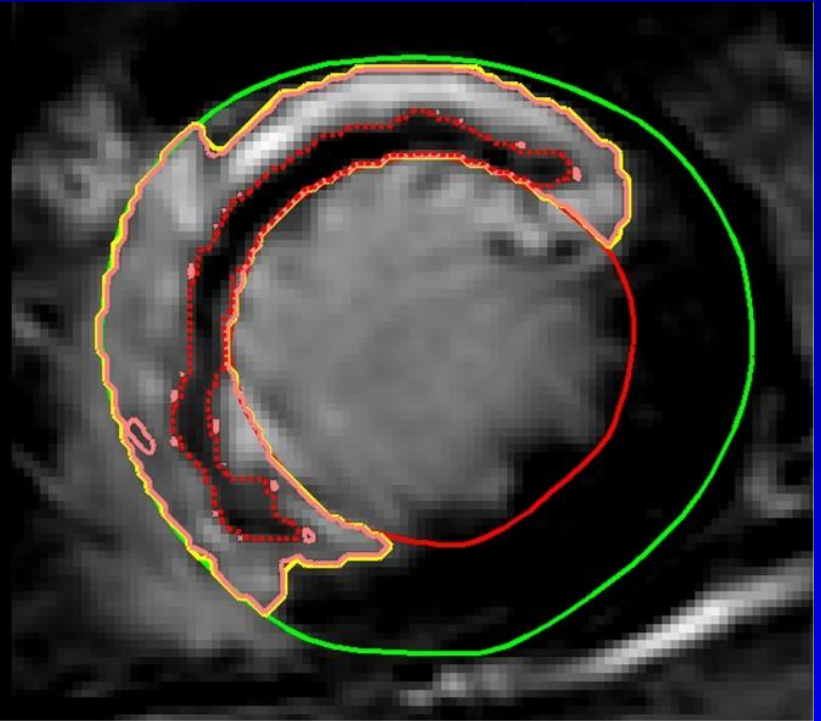


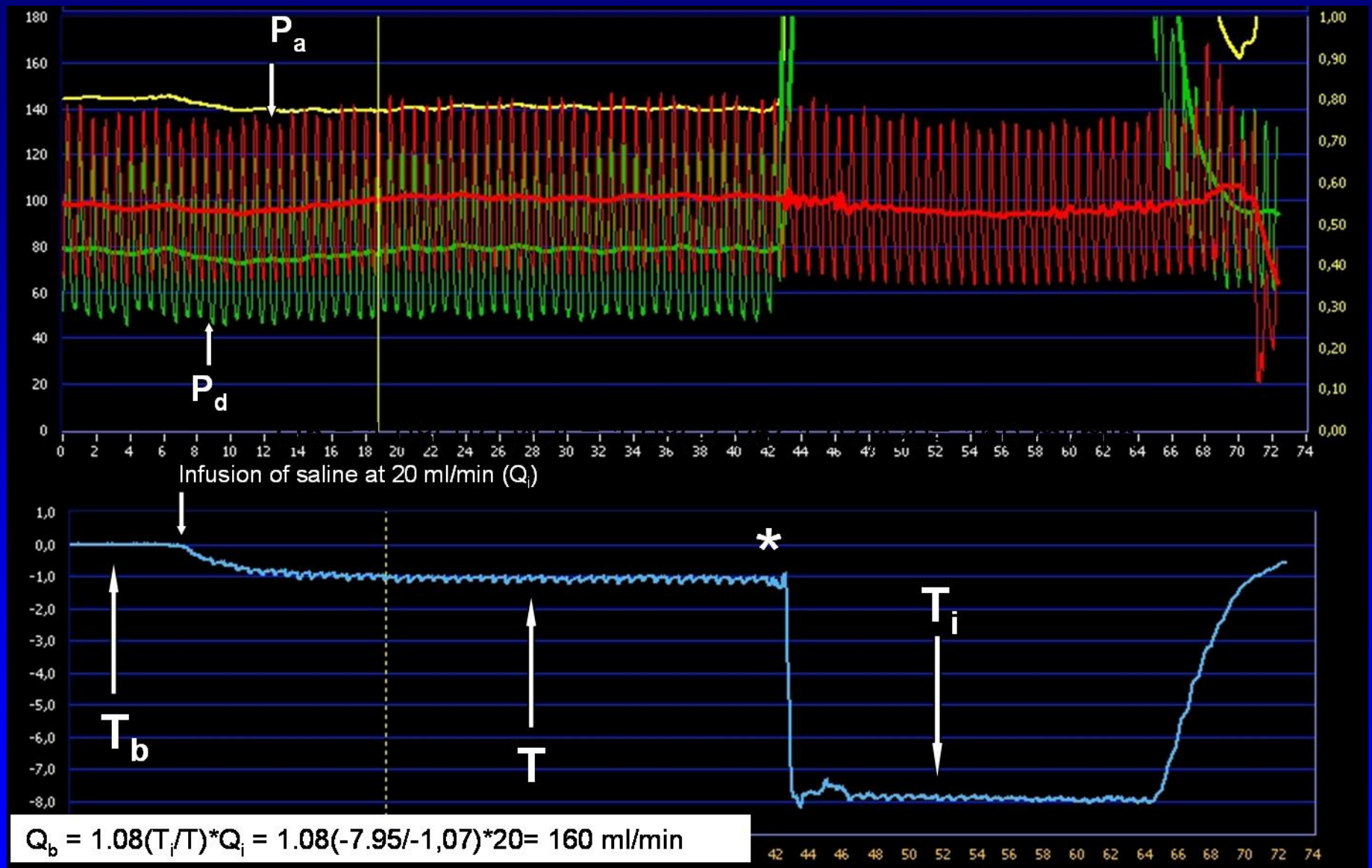
Figure 5



T2-Weighted CMR



LGE-CMR



# ***HUMAN STUDY: CONCLUSIONS (1)***

- Using this technique of continuous low rate saline infusion, *direct measurement of absolute coronary blood flow is possible*
- Within reasonable limits, the measurements are independent of infusion rate and sensor position
- Use of a *specific infusion catheter* (Hexacath, Inc, Paris) ensuring complete mixing, is paramount
- Because pressure is measured simultaneously, also coronary resistance can be calculated quantitatively
- *When  $P_w$  is also measured ( and  $FFR_{myo}$ ,  $FFR_{cor}$  and  $FFR_{coll}$  are known ) also absolute microvascular flow and resistance and absolute collateral flow and resistance can be calculated*

## NOTES:

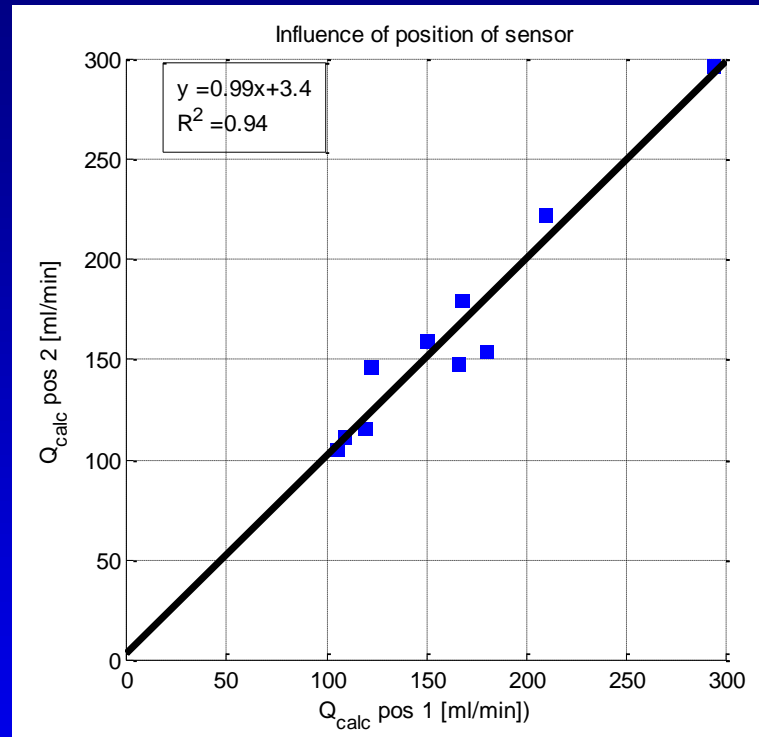
- Very reproducible, not difficult, some patience required
- Instrumentation ( introduction and connection of the infusion catheter) is not trivial and needs to be done carefully
- The infusion catheter is not commercially available yet; monorail infusion catheter is underway  
*(hexacath, Paris)*
- In my view, the method is useful for scientific purposes and very specific categories of patients, ( post-HTX, syndrome X, microvascular dysfunction, AMI) and can be used by dedicated interventionalists in the cathlab

**EINDE**

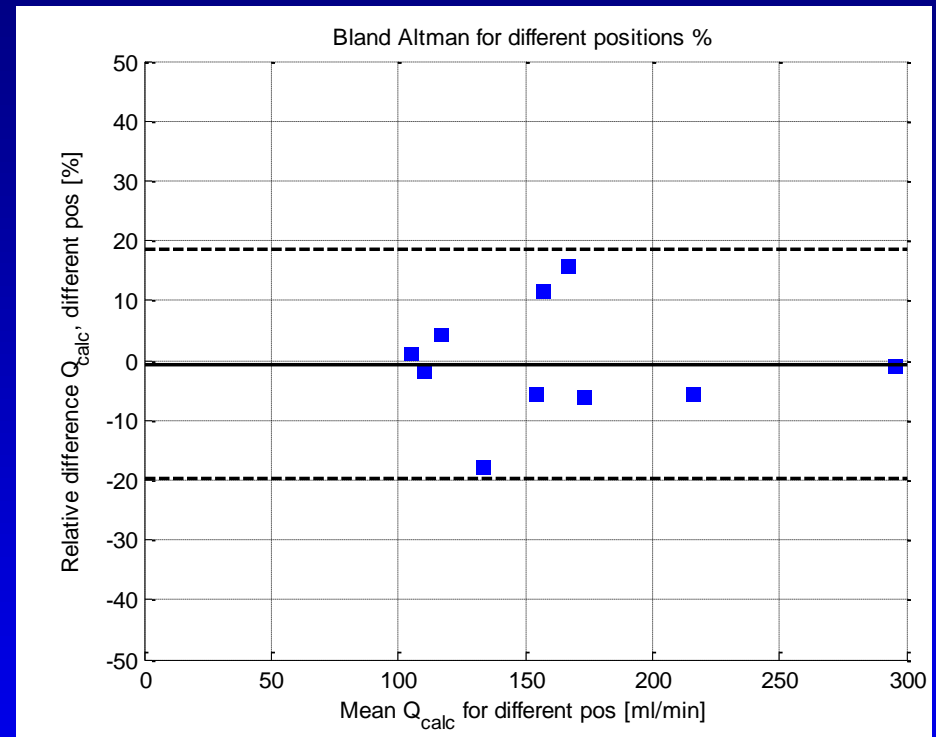


# Human study: proximal vs distal sensor position

distal position (6-8 cm)

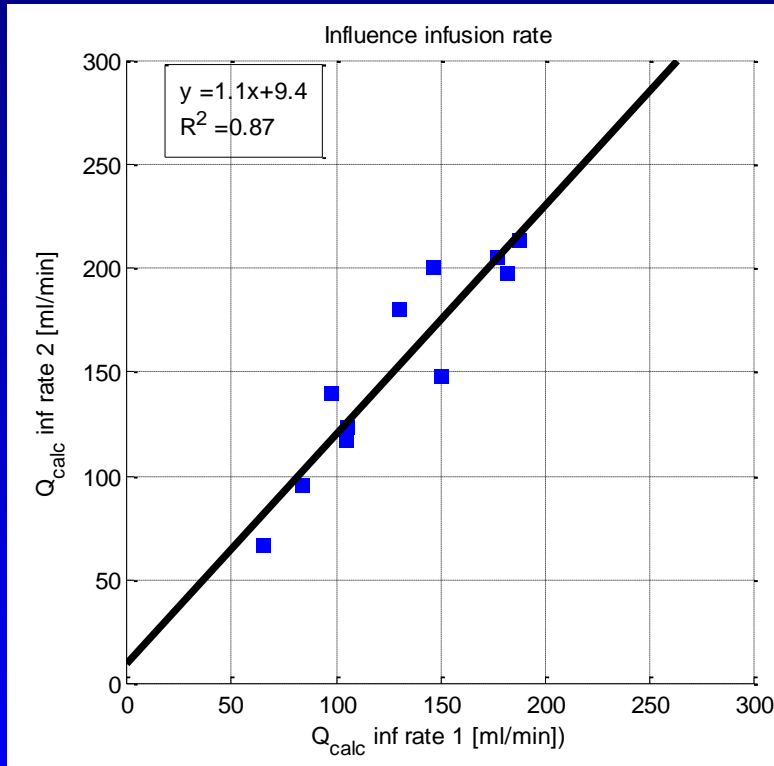


proximal position (3-4 cm)

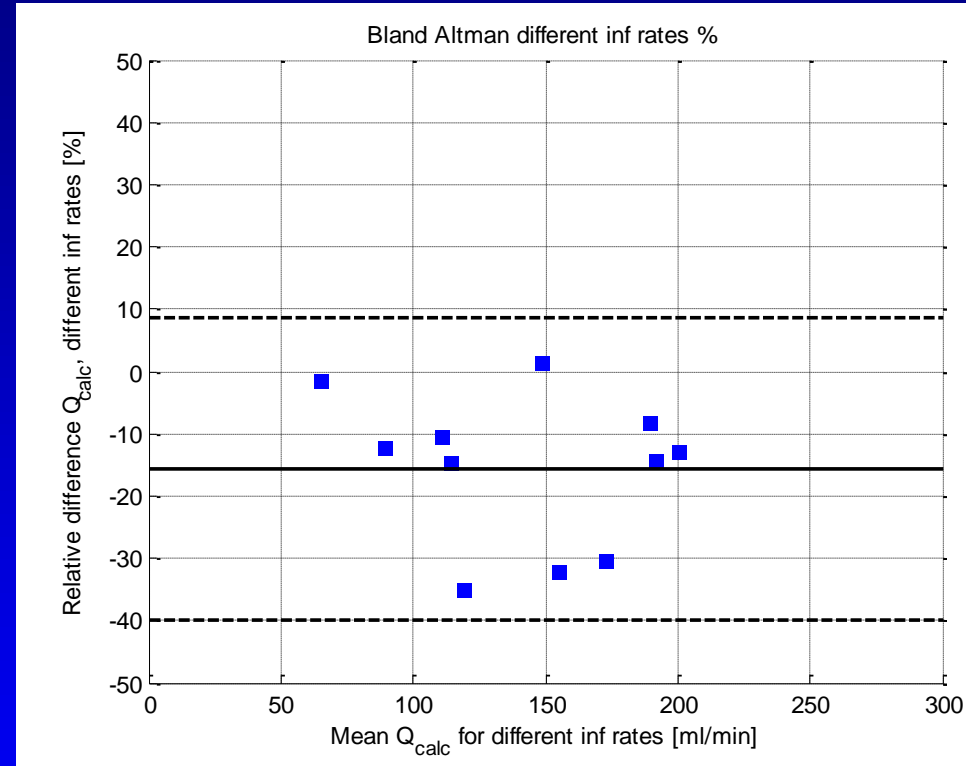


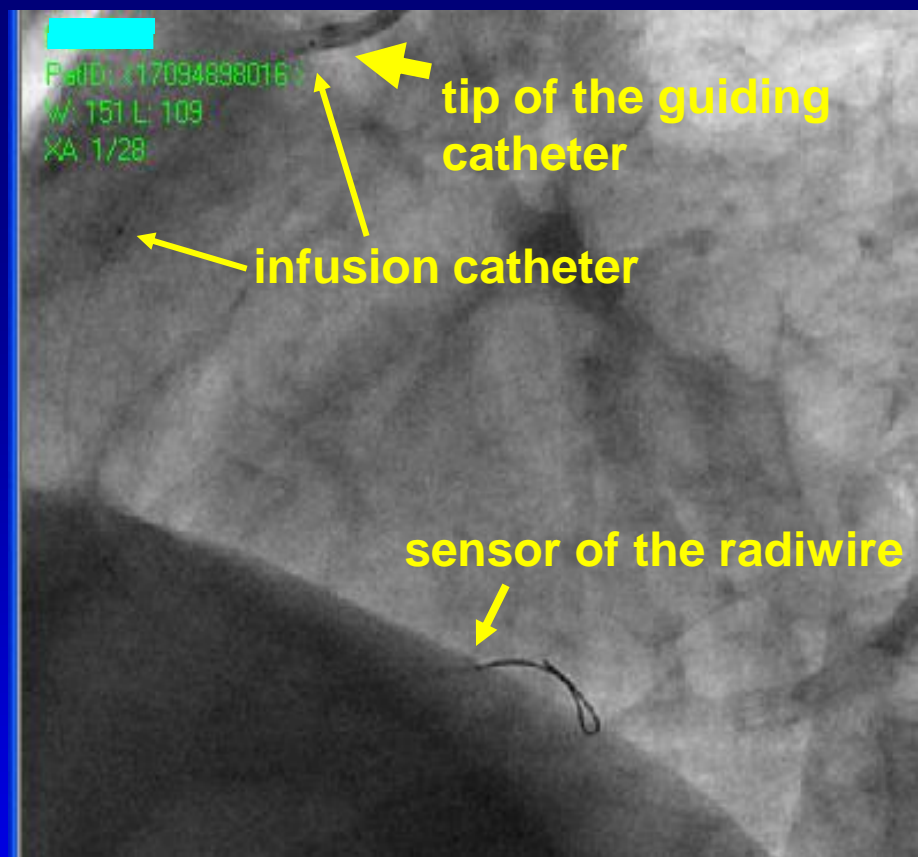
# Human study: low vs high infusion rate

high infusion rate 15- 25 ml/min

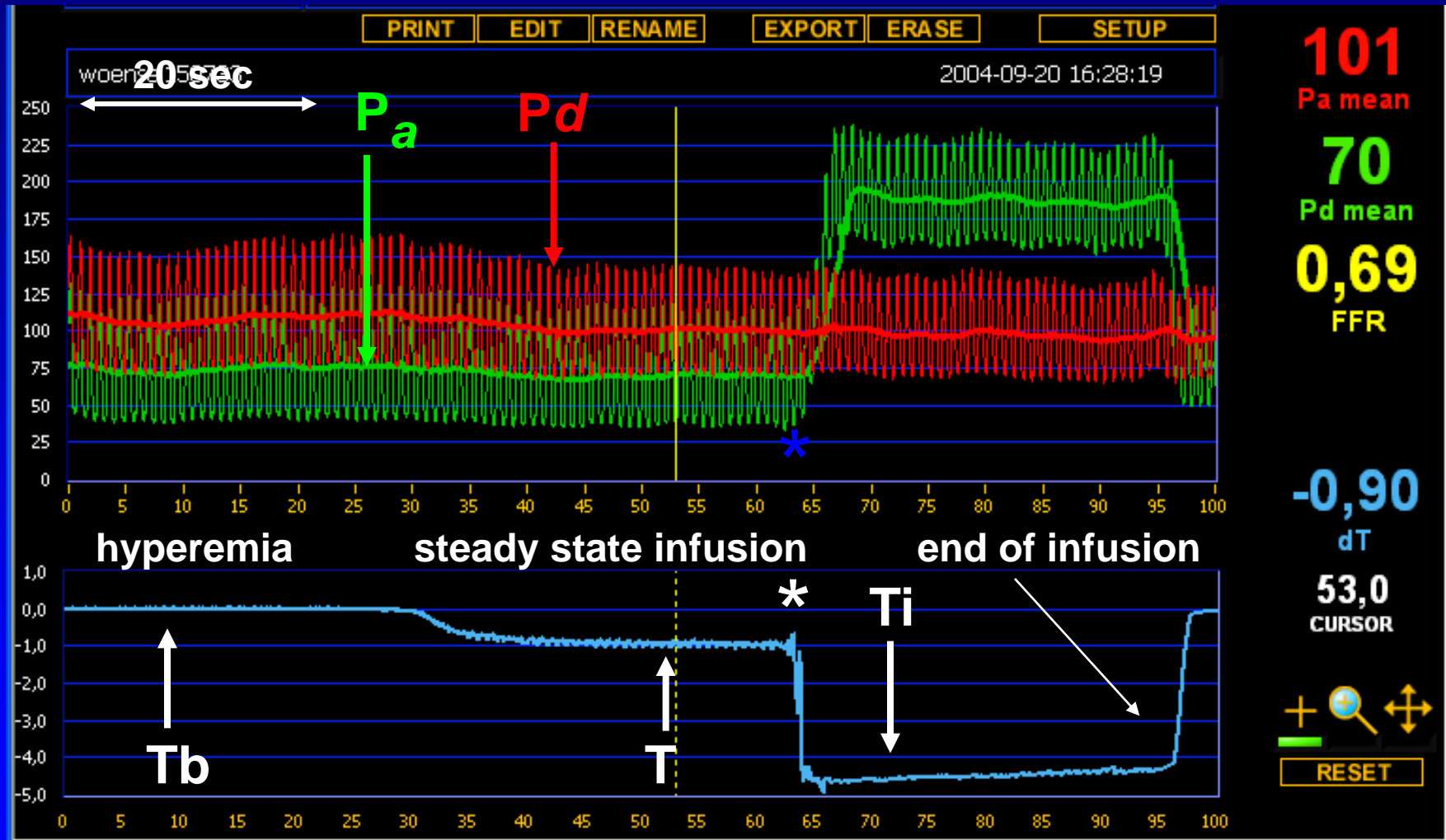


low infusion rate 10-15 ml/min





# Stenotic LAD artery, FFR = 0.69



$$Q_b = 4.5 / 0.9 * 15 * 1.08 = 81 \text{ ml/min}$$



Fladdarak

PatientID: x17094898016

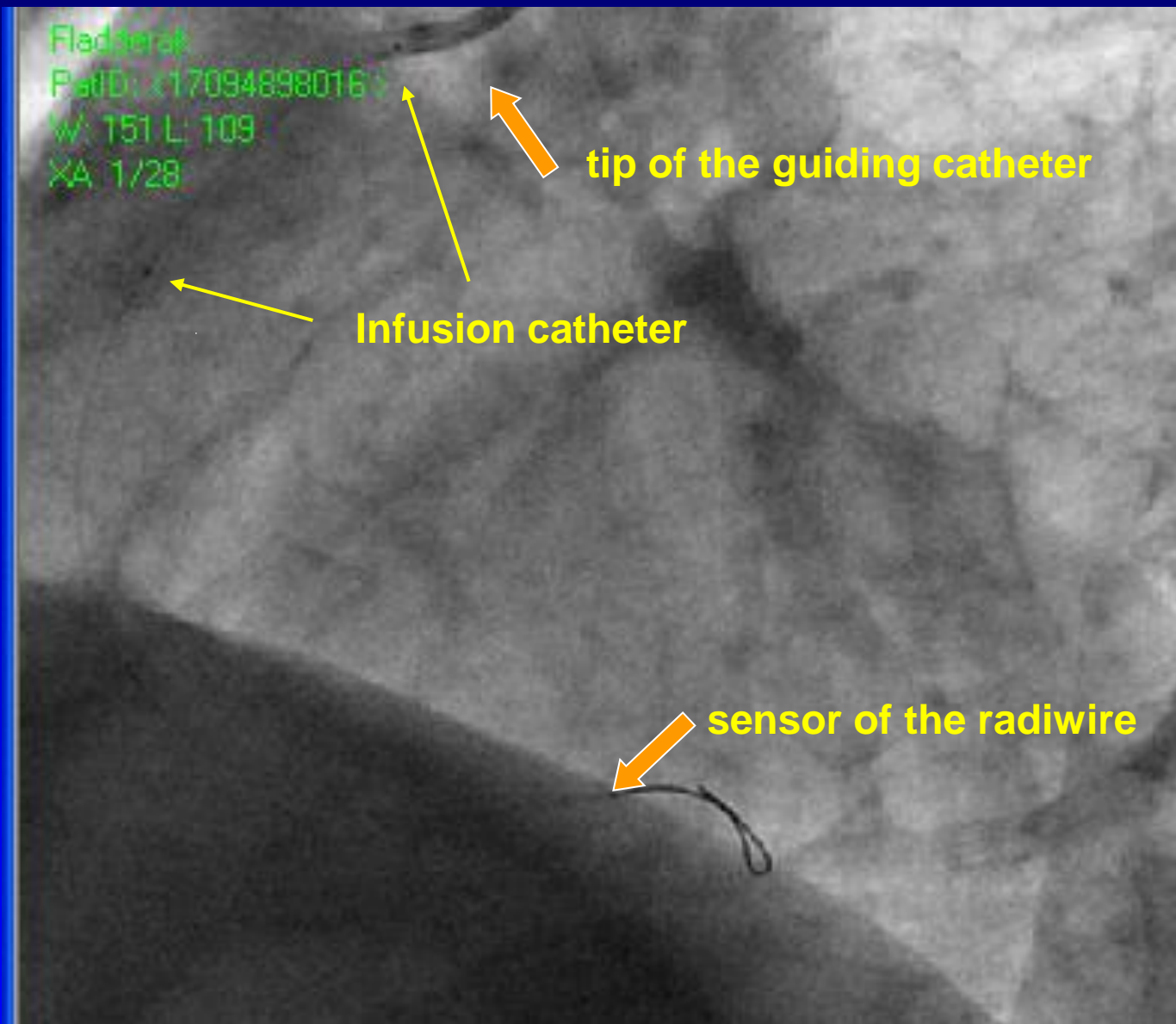
W: 151 L: 109

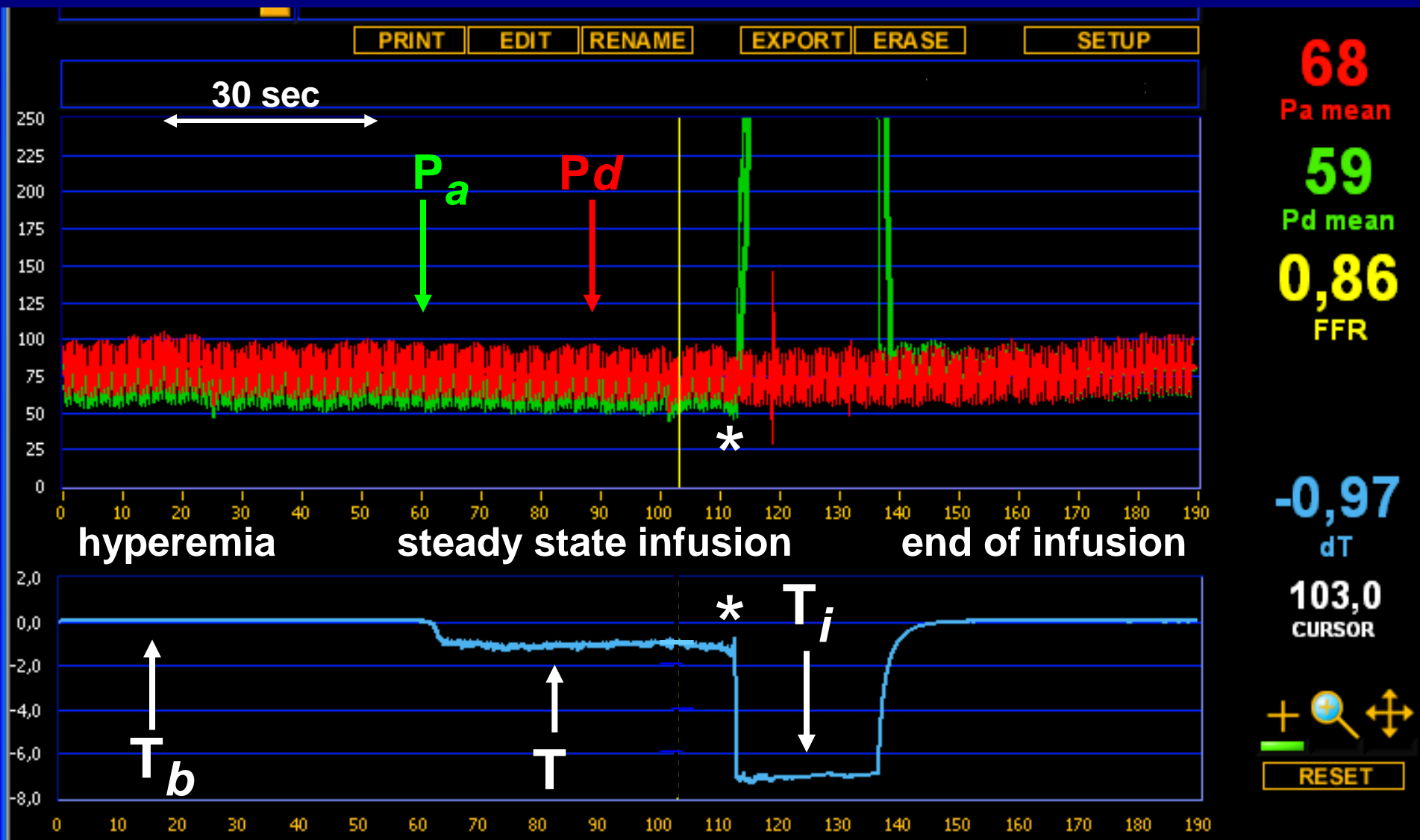
XA 1/28

tip of the guiding catheter

Infusion catheter

sensor of the radiwire





$$Q_b = 25 \times (-7.1 / -0.97) \times 1.08 = 198 \text{ ml/min}$$

*The dream of every cardio-scientist would be to know*

- **coronary, myocardial, and collateral blood flow quantitatively**

*and*

- **to be able to relate such values to the normal values for that individual patient !!**

*Together with distal coronary pressure measurement, this would also enable the calculation of*

- **true (absolute) microvascular resistance**

**enabling studying microvascular disorders, evaluation of stem-cell therapy, and many others**



## *Or vice-versa:*

- FFR after stenting is  $0.98 = 160\text{ml/min}$
- $160\text{ml/min} = 98\%$
- $164\text{ml/min} = 100\%$
- FFR before stenting was  $42\%$
- This correlates with  $164 \times 42\% = 69\text{ ml/min}$ , whereas we found  $69\text{ ml/min}$  by the first “direct” measurement

*So, for clinical decision making in the cathlab,  
absolute flow has little value.*

*Besides that, it has been impossible so far to  
measure absolute coronary or myocardial flow  
invasively*

AORTA  
100 mmHg

**pressure ( $P_d$ )**  
(mm Hg)

100

100

100

100

100

100

200

150

100

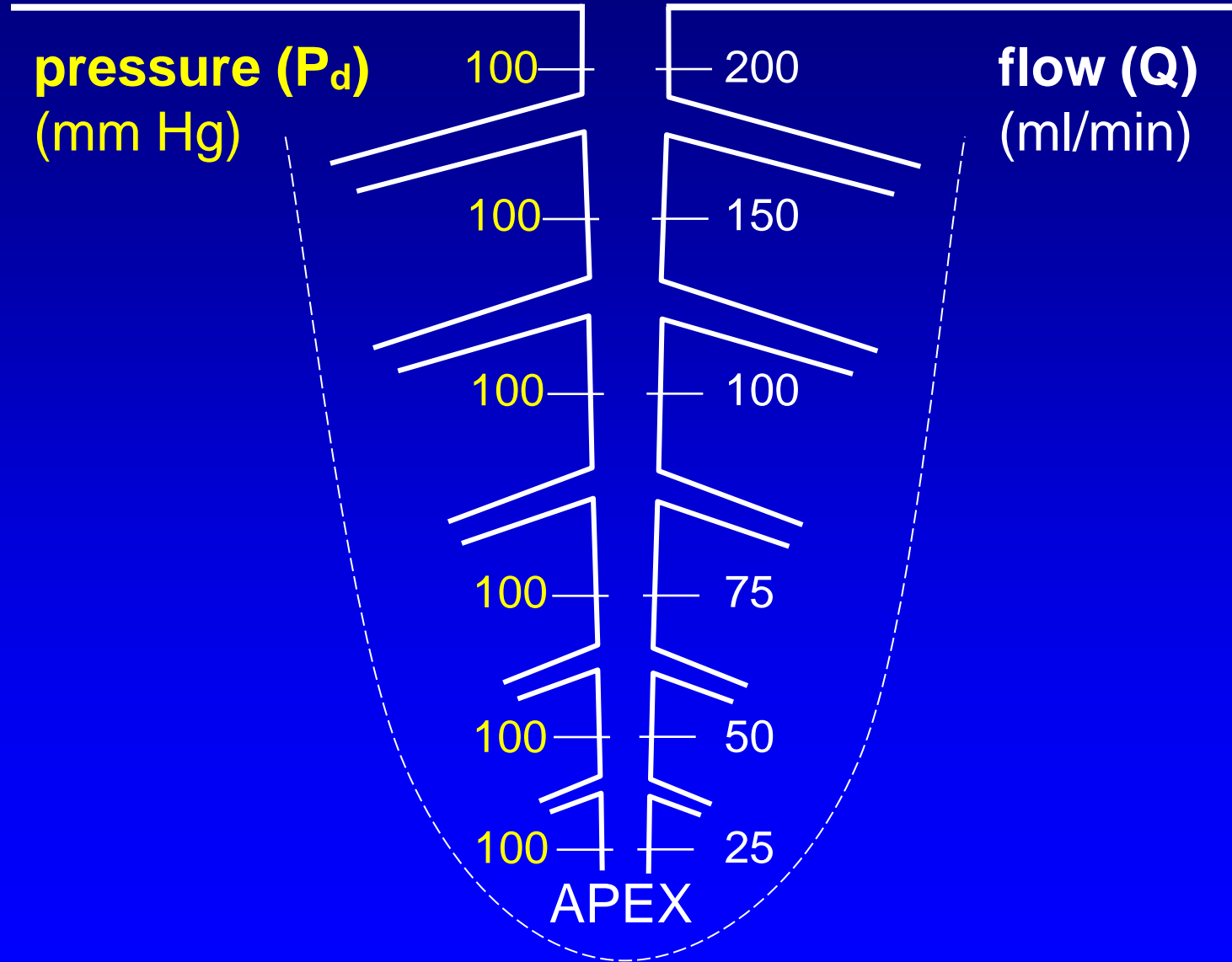
75

50

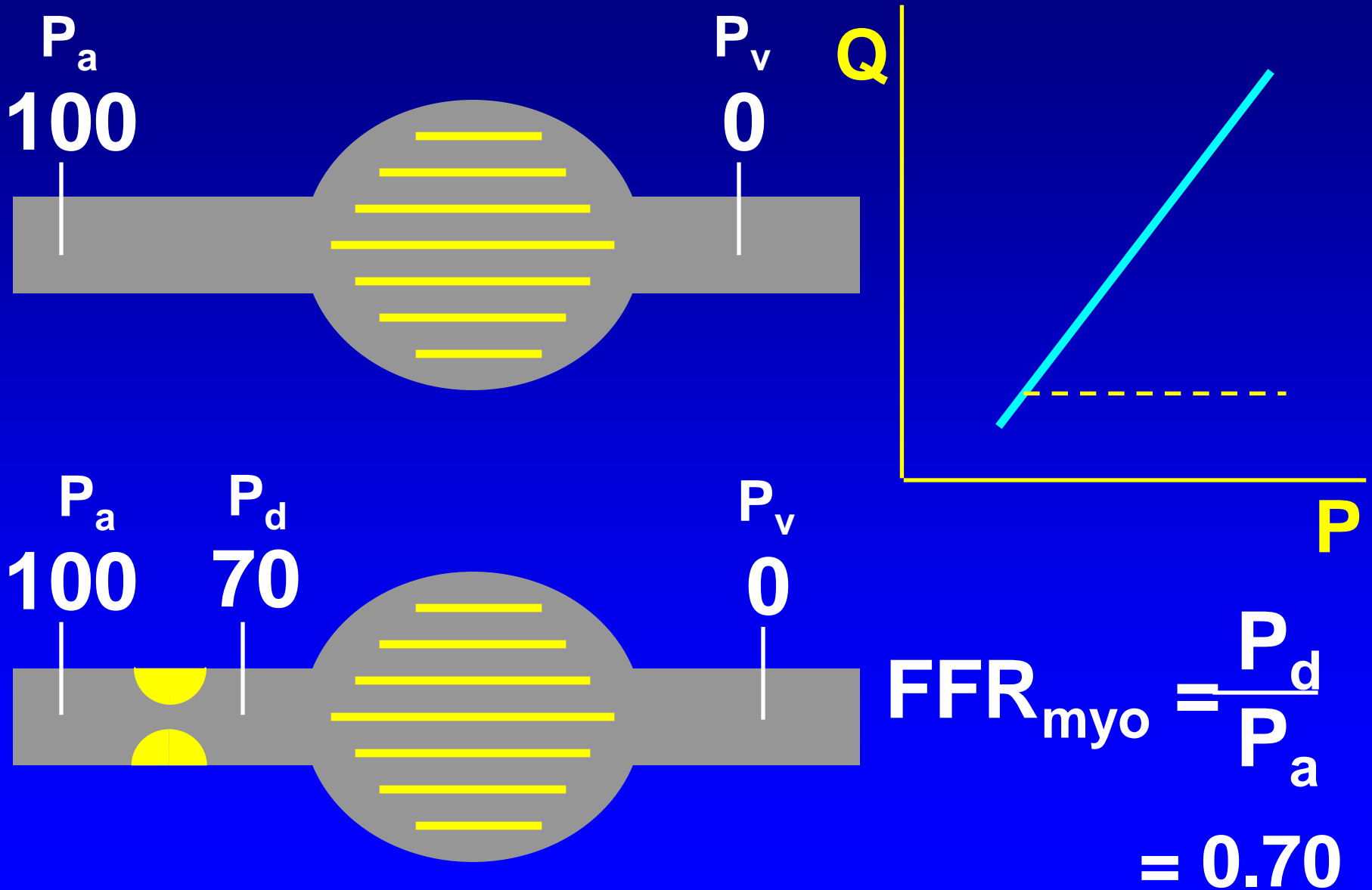
25

**flow (Q)**  
(ml/min)

APEX



# During Maximal Vasodilatation



## Pressure-flow equations:

1. Fract. Myocardial Flow Res.  $(FFR_{myo}) = \frac{P_d - P_v}{P_a - P_v}$

2. Fract. Coronary Flow Res.  $(FFR_{cor}) = \frac{P_d - P_w}{P_a - P_w}$

3. Fractional Collateral Flow  $= (FFR_{myo} - FFR_{cor})$

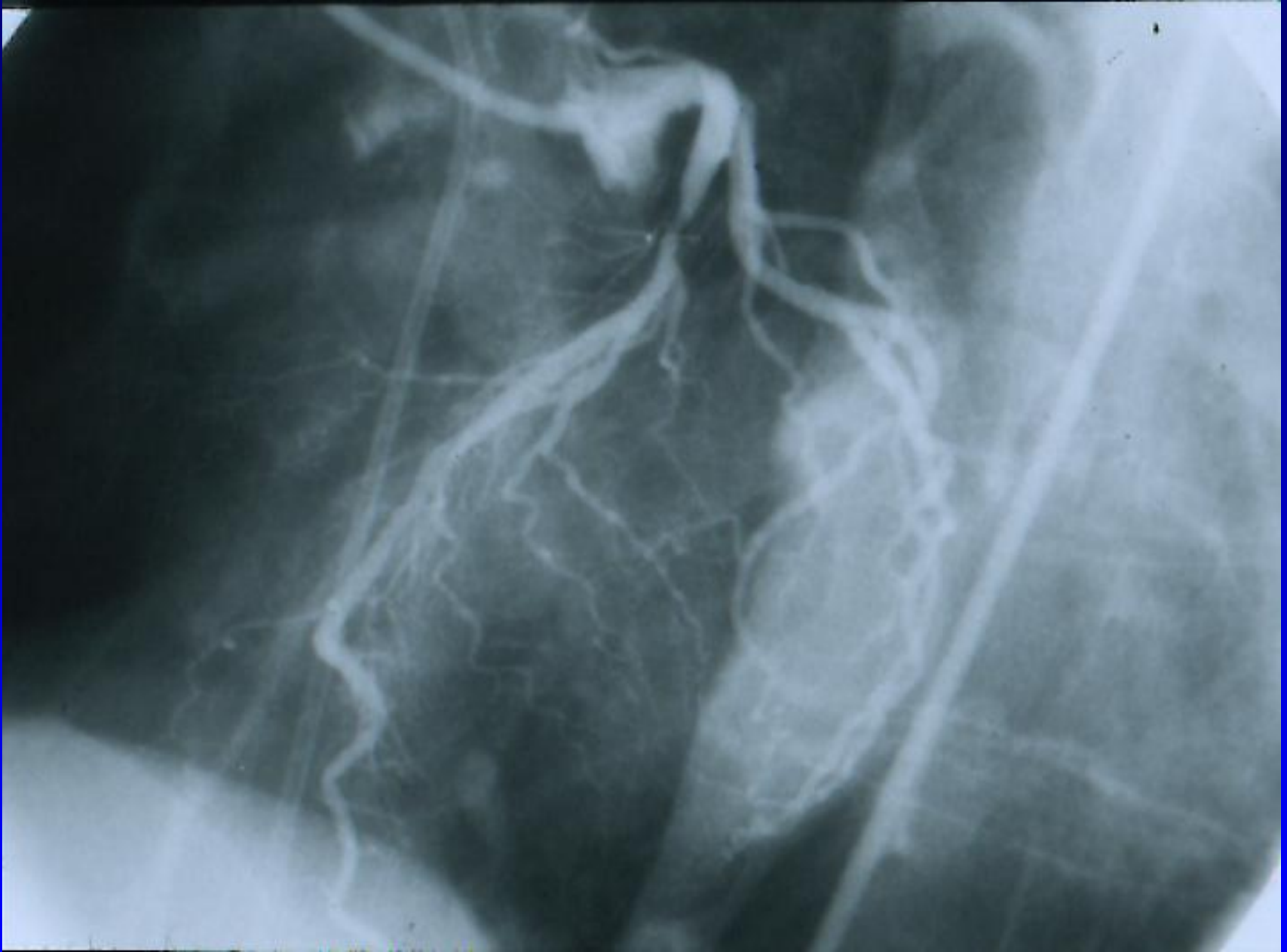
$P_a$  = mean aortic pressure at maximum hyperemia

$P_v$  = mean central venous pressure at maximum hyperemia

$P_d$  = mean distal coronary pressure at maximum hyperemia

$P_w$  = coronary wedge pressure at balloon inflation

Circulation 1993;87:1354-1367



10 vb5-PTCA - de Wit-Stek (1)



17 vb5-PTCA - de Wit-Stek (8)



## Example (1)

	before PTCA	occlusion	after PTCA
Pa	90	90	90
Pd	40	-	80
Pv	0	0	0
Pw	-	20	-
<hr/>			
FFR <sub>myo</sub> =	$\frac{40-0}{90-0}$		$\frac{80-0}{90-0}$
FFR <sub>cor</sub> =	$\frac{40-20}{90-20}$		$\frac{80-20}{90-20}$
<hr/>			
collateral flow at occlusion =	$\frac{20-0}{90-0}$		= 0.22



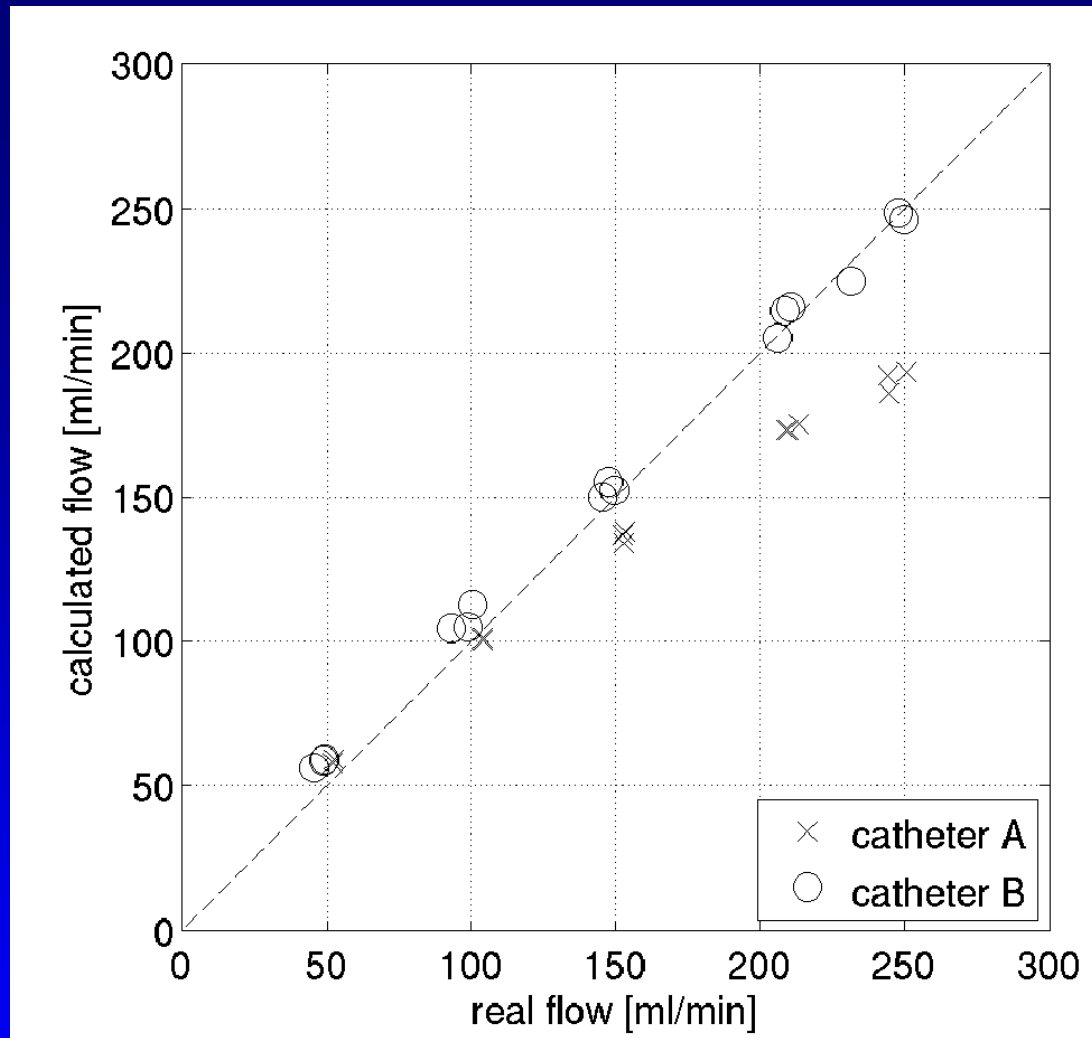


## Example (1)

	before PTCA	occlusion	after PTCA
fract. myoc. flow res.	0.44	0.22	0.89
fract. coron. flow res.	0.29	-	0.86
fract. collat. flow	0.15	0.22	0.03



***....and it would even be more attractive if absolute flow and resistance could be related to a normal value for an individual patient and individual heart rate, blood pressure, etc !***



In-vitro tests: *o = BS modified & OCCAM-2 infusion catheter*  
*x = standard infusion catheter*  
*incomplete mixing → underestimation of flow*

# ***ANIMAL STUDY: CONCLUSIONS***

- Using this technique of continuous low rate saline infusion, direct measurement of absolute coronary blood flow is possible
- Within reasonable limits, the measurements are independent of infusion rate and sensor position
- Use of a specific infusion catheter ensuring complete mixing, is paramount
- Because pressure is measured simultaneously, also resistance can be calculated quantitatively
- The method can be extrapolated to man without major modifications

FFR before stenting = 0.42

Qb before stenting = 69 ml/min

→ ***Expected normal max flow =***

$$100/42 \times 69 = \mathbf{163 \text{ ml/min}}$$

FFR before stenting = 0.42

Qb before stenting = 69 ml/min

→ *Expected normal max flow =  $100/42 \times 69 = 163$  ml/min*

FFR after stenting = 0.98

Qb after stenting = 160 ml/min

→ *“true” normal max flow =  $100/98 \times 160 = 164$  ml/min*

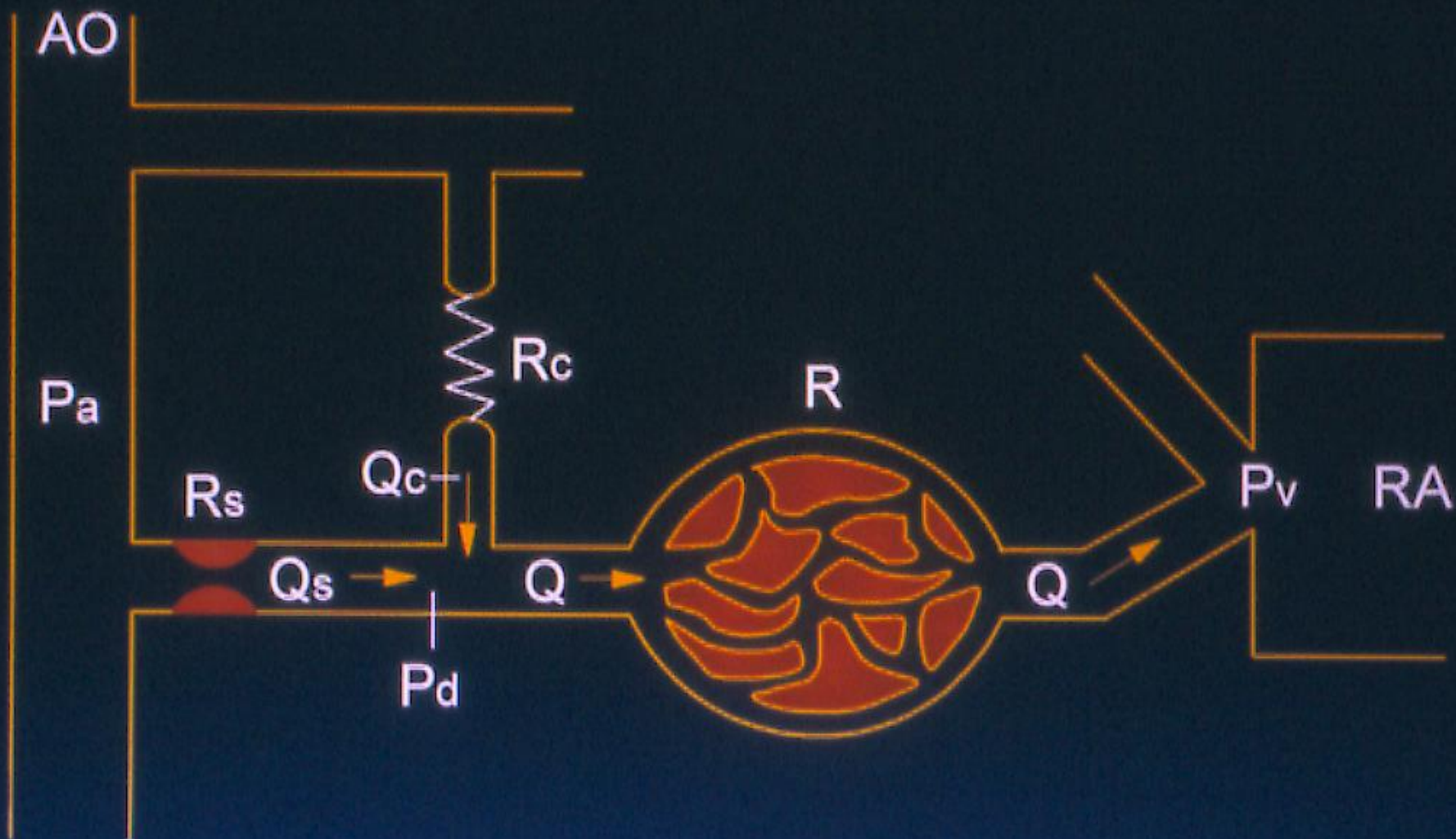
## ***Traditional coronary sinus thermodilution:***

- disappointing results, not suitable for clinical use
- not specific for one coronary artery or branch
- could not be related to a normal value
- **grossly inaccurate due to incomplete mixing:**
  - *design of infusion catheter*
  - *venous vs pulsatile arterial flow*
  - *entangling of saline along catheter*

# ***Human Study: Measurement protocol***

- introduction of pressure wire
- hyperemia by i.v. adenosine 140  $\mu\text{g/kg/min}$
- measurement of **FFR**, stop adenosine
- introduction of infusion catheter over PW
- connection of infusion catheter to infusion pump by second Y-connector
- induction of steady state hyperemia by i.v. adenosine 140  $\mu\text{g/kg/min}$
- at steady state hyperemia: zeroing of blood temperature
- start saline infusion (room temp; 10-25 ml /min =  **$Q_i$** )  
results in rapid decrease of distal blood temp by 0.5-2.0  $^{\circ}\text{C}$
- recording of steady state during 20-30 seconds (**T**)
- rapid withdrawal of PW to record infusion temperature ( **$T_i$** )
- stop adenosine; repeat sequence 2 min later





***myocardial blood flow = coronary flow + collateral flow***

**FFR myo = FFR cor + FFR coll**

**Before PCI (stenotic artery) :  $0.44 = 0.29 + 0.15$**

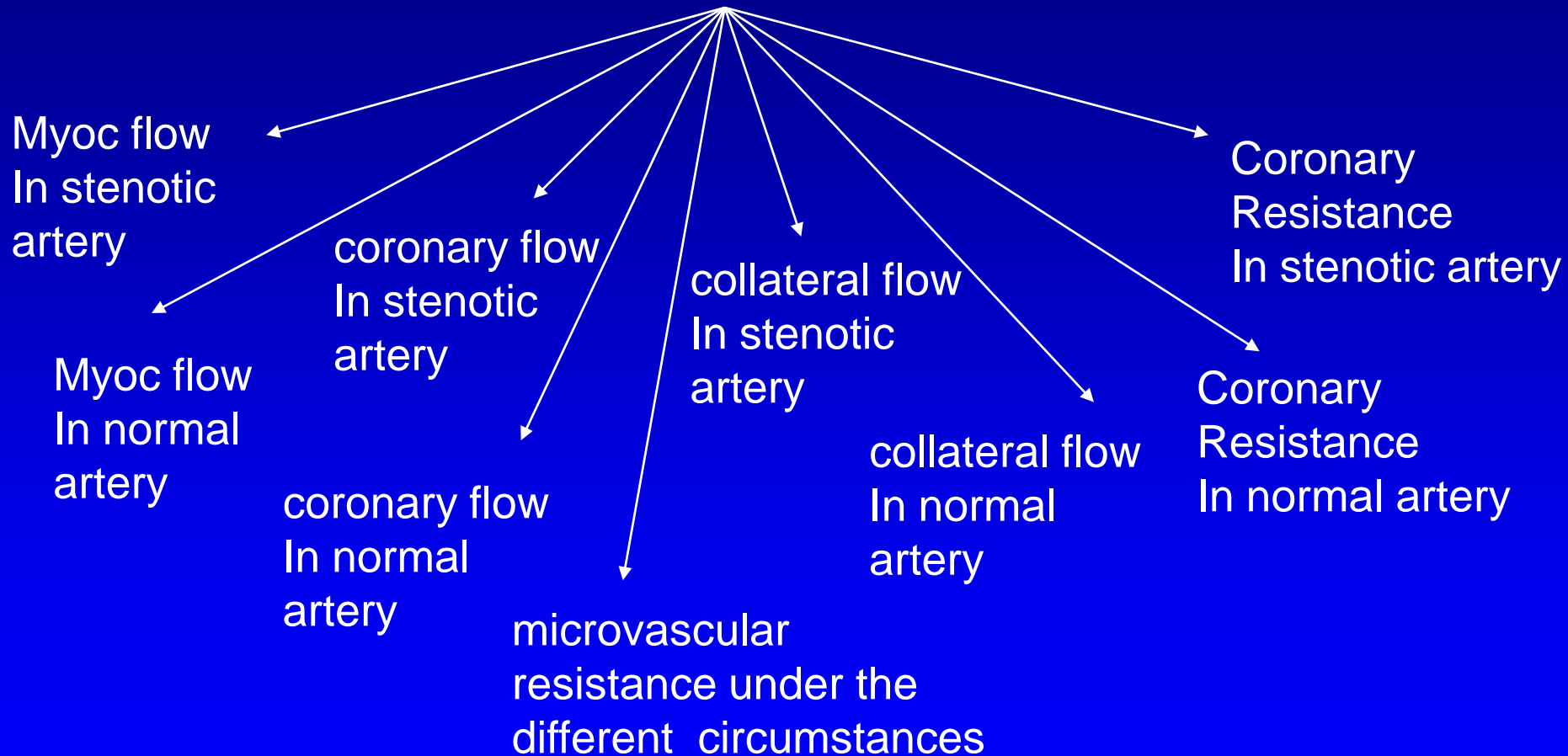
**After PCI ( stented artery) :  $0.89 = 0.86 + 0.03$**

***So, myocardial, coronary, and collateral flow both before and after PCI, are all expressed as a fraction of normal maximum myocardial blood flow***

- If one of these numbers is known quantitatively ( ml/min), all the other values are known as well**
- And if distal coronary pressure is also measured, microvascular resistance is known as an absolute number**

# FFR

*describes the mutual relation between*



# continuous infusion of saline to determine absolute maximum coronary blood flow

$$Q_b = Q_i \cdot \frac{T_i - T_b}{T - T_b} \cdot 1.08$$

*And if all temperatures are expressed relative to the  
Temperature of the blood ( zero-ed) :*

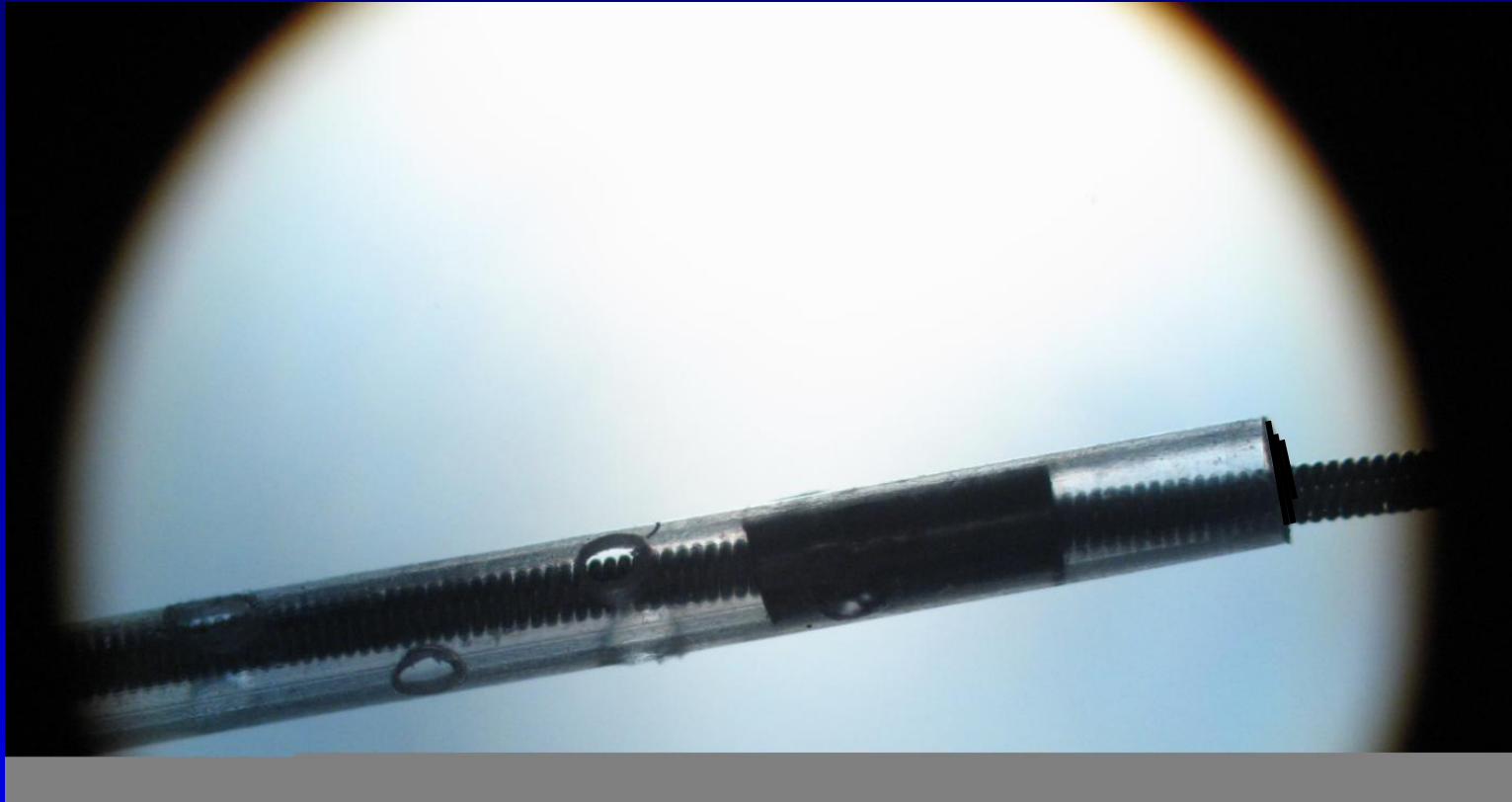
Absolute  
max coronary  
blood flow →

$$Q_b = Q_i \cdot \frac{T_i}{T} \cdot 1.08$$

infusion flow ↑

← temp of infusion at tip  
of guiding catheter

← temp of blood at  
sensor position



***OCCAM/AMT infusion catheter design:***

- 2.8 F outer diameter
- 4 sideholes in last 8 mm
- almost no infusion through tip
- maximum infusion 40 ml/min

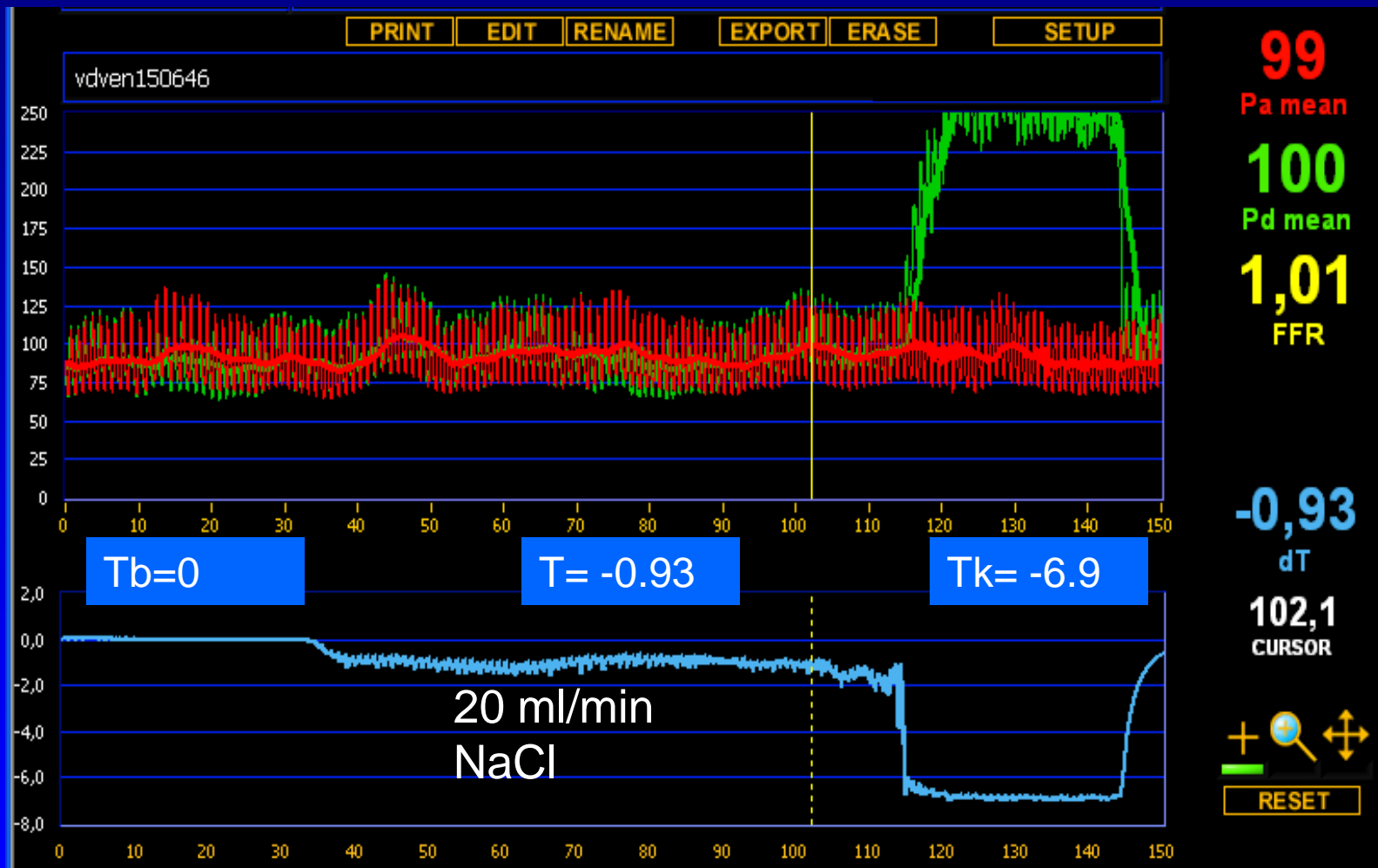
# continuous infusion of saline to determine absolute maximum coronary blood flow: theory

- Temperature sensor somewhere in the distal part of the coronary artery
- Induction of steady state hyperemia ( i.v. adenosine)
- Measurement of blood temperature (**T<sub>b</sub>**)
- Infusion of saline with known infusion rate (**Q<sub>i</sub>**) and known temperature (**T<sub>i</sub>**)
- Measurement of temperature of sensor ( **T**) after mixing of blood and infused saline
- Calculation of maximum absolute flow (**Q<sub>b</sub>**) by :

$$Q_b = Q_i \cdot \frac{T_i - T_b}{T - T_b} \cdot 1.08$$

Zierler, 1954  
Ganz, 1971

**After stenting of RCA: FFR=0.98**  
Infusion with  $Q_i = 20$  ml/min.



$$Q_b = 20 \times (6.9 / 0.83) \times 1.08 = 160 \text{ ml/min}$$